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Birth weight and blood lipid levels in Spanish adolescents: Influence of selected APOE, APOC3 and PPARgamma2 gene polymorphisms. The AVENA Study

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Published: 10 November 2008

BMC Medical Genetics 2008, 9:98 doi:10.1186/1471-2350-9-98

This article is available from: http://www.biomedcentral.com/1471-2350/9/98

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Received: 9 July 2008 Accepted: 10 November 2008

Abstract

Background: There is increasing evidence indicating that genes involved in certain metabolic processes of cardiovascular diseases may be of particular influence in people with low body weight at birth. We examined whether the apolipoprotein (APO) E, APOC3 and the peroxisome proliferator-activated receptor- γ -2 (PPAR γ 2) polymorphisms influence the association between low birth weight and blood lipid levels in healthy adolescents aged 13–18.5 years.

Methods: A cross-sectional study of 502 Spanish adolescents born at term was conducted. Total (TC) and high density lipoprotein cholesterol (HDLc), triglycerides (TG), apolipoprotein (apo) A and B, and lipoprotein(a) [Lp(a)] were measured. Low density lipoprotein cholesterol (LDLc), TC-HDLc, TC/HDLc and apoB/apoA were calculated.

Results: Low birth weight was associated with higher levels of TC, LDLc, apoB, Lp(a), TC-HDLc, TC/HDLc and apoB/ apoA in males with the APOE $\varepsilon 3\varepsilon 4$ genotype, whereas in females, it was associated with lower HDLc and higher TG levels. In males with the APOC3 S1/S2 genotype, low birth weight was associated with lower apoA and higher Lp(a), yet this association was not observed in females. There were no associations between low birth weight and blood lipids in any of the PPAR $\gamma 2$ genotypes.

Conclusion: The results indicate that low birth weight has a deleterious influence on lipid profile particularly in adolescents with the APOE $\varepsilon 3/\varepsilon 4$ genotype. These findings suggest that intrauterine environment interact with the genetic background affecting the lipid profile in later life.

Background

There is increasing evidence indicating that low birth weight is a risk factor for cardiovascular disease. Indeed, among persons born at term, those with lower birth weights have a higher relative risk of metabolic health problems in early adulthood, including hypertension, cardiovascular disease, and type 2 diabetes [1]. In this context, findings from the AVENA Study (*Alimentación y Valoración del Estado Nutricional en Adolescentes*, Food and Nutritional Status in Spanish Adolescents) indicated that birth weight is associated with a number of cardiovascular disease risk factors in adolescents such as total and central body fat, fat free mass, and muscular fitness [2-5].

One of the hypotheses explaining this phenomenon is the predisposition to adopt a "thrifty phenotype". According to this theory, fetal adaptation to an adverse intrauterine environment involves programming of pathways that might predispose to metabolic abnormalities and cardiovascular disease in later life. The intra-uterine under- or over-nutrition can alter the gene expression of the fetus, causing developmental adaptations that may lead to permanent changes in physiology and metabolism; changes that may have consequences later in life.

Findings of several cohort studies indicate that low birth weight is associated with adverse plasma lipid profile [6-8], whereas others have suggested that this association is too small to be considered of public health importance and that other variables as postnatal weight gain could act as confounder of this relationship [9]. One would predict that genes involved in the metabolic processes of these diseases may have different effects on people with different body weight at birth. This phenomenon has been described in relation to blood lipids and the apolipoprotein (APO) E gene [10], and both blood lipids and insulin resistance and the peroxisome proliferator-activated receptor- γ -2 (PPAR γ 2) gene [11,12]. We have studied the effect of the Ala12 allele in the PPARy2 gene on the relationship between birth weight and body composition in Spanish adolescents, and we showed that small body weight at birth may program lower fat free mass in adolescents carrying the Ala12 allele [13].

Coronary heart disease is a leading cause of global mortality. The relationship between blood lipids and the development of coronary heart disease in children and adolescents is well established. Results from the AVENA Study showed that 20–30% of Spanish adolescents present an unfavorable blood lipid profile [14,15]. In the AVENA study, the APOE, APOC3 and PPARy2 gene polymorphisms were genotyped. The ApoE gene is one of the most important genetic determinants of atherogenesis, since its major function is to regulate the hepatic clearance of triglycerides (TG) rich particles such as chylomicron remnants and very low density lipoprotein remnants. The APOC3 gene have also been proposed as being potentially responsible for the occurrence of lipid profile disturbances, since it has an inhibitory effect on lipoprotein lipase activity and hepatic uptake of lipoproteins [16]. Finally, the PPAR γ -2 has been shown to enhance lipoprotein TG hydrolysis by endothelial lipoprotein lipase, which in turn seems to affect lipid profile [17].

To understand whether an adverse intrauterine environment may alter the expression of these cardiovascular disease-related genes is clinically relevant. This study aims to clarify to what extent variants of these genes might interact with birth weight in determining the blood lipid profile later in life. Therefore, we examined the influence of APOE, APOC3 and PPAR γ 2 gene polymorphisms on the association between low birth weight and blood lipid levels in Spanish adolescents from the AVENA Study.

Methods

The AVENA Study is a cross-sectional study designed to assess the nutritional status of a representative sample of Spanish adolescents aged 13 to 18.5 years. Data collection took place from 2000 to 2002 in five Spanish cities (Madrid, Murcia, Granada, Santander and Zaragoza). The complete methodology of the study has been described in detail elsewhere [15,18]. The number of adolescents included in the AVENA Study was 2859 adolescents. Blood samples and DNA data were randomly obtained from 502 participants. The subgroup from which blood samples were obtained was similar to the remaining subjects regarding the variable selected to calculate the number of participants to be included in the study, i.e. body mass index (BMI) [15], as well as regarding age (P = 0.750) and gender proportions (P = 0.320). A comprehensive verbal description of the nature and purpose of the study was given to the adolescents, their parents and teachers. Written consent to participate was requested from both parents and adolescents. Adolescents with personal history of cardiovascular disease, under medication at the time of the study, or those who were pregnant, were excluded. The study protocol was performed in accordance with the ethical standards laid down in the 1961 Declaration of Helsinki (as revised in Hong-Kong in 1989, and in Edinburgh in 2000), and approved by the Review Committee for Research Involving Human Subjects of the Hospital Universitario Marqués de Valdecilla (Santander, Spain).

Before any testing was performed, the parents completed a questionnaire, part of which addressed the adolescents' previous and current health status. Socioeconomic status was also assessed via the questionnaire, and was defined by the educational level and occupation of the father. According to this information, and following the recommendation of the Spanish Society for Epidemiology, the adolescents were classified into five categories: low (I), medium-low (II), medium (III), medium-high (IV) and high socioeconomic status (V).

Neonatal data

Data on birth weight and gestational age at birth were obtained from health booklets records that are issued at birth and where child's paediatricians records birth weight, charts the infant's growth and vaccinations. Birth weight was expressed as the standard deviation from the expected weight calculated with the use of reference standards previously described for this population, according to sex and gestational age [19]. This variable (called birth weight score) will be used for the analyses.

Gestational age was coded as 1 for those who were born between the 35th to 40th week of gestation, and 2 for those who were born after the 40th week of gestation. According to this information, 80.9% and 77.0% of males and females, respectively, were born between 35 to 40 weeks of gestation, and 12.2% of males and 15.1% of females were born after more than 40 weeks of gestation. The percentage of adolescents born before 35 weeks of gestation was 6.9% of the males and 7.9% of the females. This group was not included in the analyses.

Physical examination

Anthropometric measurements were obtained as described elsewhere [20,21]. BMI was calculated as weight in kilograms divided by square of height in meters (kg/m²). Skinfold thickness was measured at the biceps, triceps, subscapular, suprailiac, thigh and calf on the left side of the body to the nearest 0.2 mm using a Holtain skinfold caliper. Body fat percentage was calculated from skinfold thicknesses (triceps and subscapular) using Slaughter's equations [22], and fat free mass (kg) was derived by subtracting fat mass from total body weight. Reference values for anthropometric measurements of the AVENA Study can be found elsewhere [20,21].

Identification of pubertal development was assessed according to Tanner & Whitehouse [23]. Self-reported genital development in males and breast development in females were used for pubertal stage classification.

Cardiorespiratory fitness

Cardiorespiratory fitness was assessed by the 20 m shuttle run test as previously described [24]. It was considered as the number of stages completed (precision of 0.5 steps). All participants were familiarized with the test, since the 20 m shuttle run test is one of the fitness tests included in the physical education curriculum in our country. Reference values of fitness levels have been reported for the whole study population as well as for those adolescents from whom blood sample was obtained [25].

Blood sampling

Blood (20 ml) was collected from an antecubital vein between 8:00 and 9:00 a.m, after an overnight fast. Serum levels of total (TC) and high density lipoprotein cholesterol (HDLc), as well as the levels of TG, apolipoprotein (apo)A and B, and lipoprotein(a) [Lp(a)] were measured. The coefficients of variation were less than 3% and the intra-class coefficients were higher than 0.96% for all blood variables. Quality control of the assays was assured by the Regional Health Authority, as is compulsory for all hospital clinical laboratories in Spain.

Low density lipoprotein cholesterol (LDLc) was calculated with the Friedewald formula [26]. The following atherogenic indices were also calculated: TC-HDLc, TC/ HDLc, and apoB/apoA. A detailed description of the blood analysis as well as reference values for lipid and lipoprotein has been reported by Ruiz et al. [15,27].

Genotyping

Genomic DNA was extracted and purified from 500 μ L of whole blood treated with EDTA, using the Quiagen procedure described by Higuchi [28]. APOE genotypes were determined by polymerase chain reaction (PCR) and allele-specific restriction digestion of the amplified products with the restriction enzyme Hhal, as described elsewhere [29] (rs7412, and rs429358). APOE genotypes were encoded as $1 = \epsilon 2/\epsilon 3$, $2 = \epsilon 3/\epsilon 3$, and $3 = \epsilon 3/\epsilon 4$. APOC3 genotypes were determined by PCR and allele-specific restriction digestion of the amplified products with the restriction enzyme SstI following the procedures described elsewhere [30]. APOC3 genotypes were encoded as 1 = S1/S1 and 2 = S1/S2 (rs5128). The PPARy2 genotypes were determined by the PCR method and further digestion of products with BstU-I restriction enzymes as previously described [31], and were encoded as 1 = Pro12/Pro, and 2 = Pro12/Ala and Ala12/Ala genotype [32] (rs1801282). The percentage of adolescents with the Ala12/Ala genotype was 1.3%, therefore this sub-group was analyzed together with the Pro12/Ala.

Statistical analysis

Following a bivariate correlation analysis, multiple regressions were used to study the association between birth weight and blood lipid levels, for males and females separately (unadjusted model). Further analyses were done after controlling for age, pubertal stage, socioeconomic status, gestational age, BMI and cardiorespiratory fitness (adjusted model). A separate regression model was performed for each lipid parameter. Additional analyses were also performed by genotypes. To stabilize variability and to achieve normality in the residuals, TC, HDLc, LDLc, TG, apoA, apoB and Lp(a) were transformed to the natural logarithm.

The impact on blood lipid levels of APOE, APOC3 and PPARy2 genotypes was analyzed by one-way analysis of variance (ANOVA), for males and females separately. For the APOE genotypes, the subgroup means were compared by Tukey's test. Comparisons were adjusted for mass significance as described by Holm [33,34]. The method of Holm proceeds as follows: Sort the P-values of the k tests in increasing order, P_1 , P_2 , ..., P_k . If $P_1 > \alpha/k$; none of the k tests are significant, and the test procedure is finished. If $P_1 \le \alpha/k$, test 1 is significant, and now P_2 is examined. If $P_2 > \alpha/(k-1)$, none of the (k-1) remaining tests are significant, but if $P_2 \le \alpha/(k-1)$, test 2 is significant and P_3 is examined. This procedure goes on until $P_i > \alpha/(k - i + i)$ 1), and the procedure is interrupted. This method keeps family error rate less than α . Family error rate is defined as the probability that one or more false significances out of k tests is less than or equal to α .

Interaction effects between birth weight and sex, and between birth weight and genotypes were tested by inserting product terms for the relevant variables. The analyses were performed using the Statistical Package for Social Sciences (SPSS, v. 15.0 for WINDOWS; SPSS Inc, Chicago) and the level of significance was set to 0.05.

Results

Descriptive characteristics of the study sample are shown in Table 1.

Interactions

There was a significant (all P < 0.05) interaction effect between birth weight and sex in all the outcome variables, therefore, all the analyses were performed separately for males and females. There was a significant interaction effect between birth weight and APOE in all of the outcome variables (TC, P = 0.02; HDL, P = 0.021; LDLc, P = 0.001; apoA, P = 0.049; apoB, P = 0.01; Lp(a), P = 0.051; TC-HDLc, P = 0.025; apoB/apoA, P = 0.054), except for TG (P = 0.362). There was also a significant interaction effect between birth weight and APOC3 (for TG, P = 0.025; apoA, P = 0.038, and Lp(a), P = 0.002). No significant (all P > 0.1) interaction effect between birth weight and PPARy2 was found.

Birth weight and lipids

In males, low birth weight was associated with lower levels of HDLc and apoA, and higher TC/HDLc and apoB/ apoA ratios (Figure 1). Birth weight was not significantly associated with blood lipids in females. The results were similar after controlling for age, pubertal stage, socioeconomic status, gestational age, BMI and cardiorespiratory fitness (Table 2).

Table I: Descriptive characteristics of the study sample

	Males (n = 260)	Females (n = 242)
Age (years)	15.4 ± 1.4	5.4 ± .4
Tanner II,III,IV,V (%)	5.8, 15.1, 41.8, 37.3	1.6, 8.1, 54.3, 36.0
Weight (kg)	64.6 ± 13.4	56.8 ± 10.5
Height (cm)	170.7 ± 8.2	161.5 ± 6.4
Body mass index (kg/m ²)	22.1 ± 3.9	21.7 ± 3.5
Cardiorespiratory fitness (stage)	7.0 ± 2.6	4.1 ± 1.8
Sum of 6 skinfolds	75.8 ± 38.1	98.8 ± 31.9
Percentage body fat	20.2 ± 10.4	25.6 ± 6.6
Triglycerides (mg/dl)	71.3 ± 31.6	64.7 ± 26.4
TC (mg/dl)	156.7 ± 26.4	169.9 ± 26.4
HDLc (mg/dl)	51.0 ± 9.7	59.2 ± 11.6
-DLc (mg/dl)	91.4 ± 23.9	97.4 ± 23.2
ApoA (mg/dl)	116.9 ± 17.1	127.7 ± 19.2
ApoB (mg/dl)	66.9 ± 14.3	69.9 ± 13.8
_p(a) (mg/dl)*	14.1 ± 4.2	5.3 ± 3.9
ApoA/ApoB (mg/dl)	1.8 ± 0.5	1.9 ± 0.5
TC-HDLc (mg/dl)	105.7 ± 25.9	110.6 ± 25.8
Birth weight (g)	3443.0 ± 539.0	3283 ± 564
Birth weight score	409.0 ± 516.0	353 ± 559
Socieconomic status I,II,III,IV,V (%)	5.9, 25.3, 41.6, 23.6, 3.6	7.2, 25.4, 37.1, 23.3, 6.9

Values are means \pm SD, otherwise indicated.

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo, apolipoprotein; Lp(a), lipoprotein a.

*Geometric mean \pm SD.

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Table 2: Regression coefficients (eta) showing the association
between birth weight score and blood lipid and lipoprotein levels
by sex after controlling for age, pubertal stage, socioeconomic
status, gestational age, body mass index and cardiorespiratory
fitness

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	Males	(n = 260)	Females (n = 242)		
	\hat{eta}	P value	\hat{eta}	P value	
TC*	0.035	0.682	0.073	0.476	
HDLc*	0.134	0.009	0.058	0.537	
LDLc*	-0.05 I	0.549	-0.044	0.668	
TG*	-0.017	0.841	-0.06	0.541	
apoA*	0.187	<0.001	-0.025	0.784	
apoB*	-0.03 I	0.723	0.032	0.749	
Lp(a)*	-0.085	0.334	0.051	0.614	
TC-HDLc	-0.026	0.76	0.044	0.668	
TC/HDLc	-0.066	0.003	-0.001	0.991	
ароВ/ароА	-0.113	0.029	0.049	0.609	

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo, apolipoprotein; Lp(a), lipoprotein a. *Analyses were performed on log transformed data.

Genotypes and lipids

All the genotype frequencies of the analyzed gene polymorphism were in agreement with the Hardy-Weinberg equilibrium. The distribution of the APOE genotypes did not differ between males and females (P = 0.961) (Table 3). Males with the $\epsilon 2\epsilon 3$ genotype had lower levels of TC, LDLc and apoB compared with those with the $\epsilon 3\epsilon 3$ genotype. Similarly, females with the $\epsilon 2\epsilon 3$ genotype had lower levels of LDLc, and apoB compared with those with the $\epsilon 3\epsilon 3$ genotype. Females with $\epsilon 2\epsilon 3$ genotype had higher levels of apoA compared with those with the $\epsilon 3\epsilon 3$ genotype. There were no subjects with the $\epsilon 2\epsilon 2$, $\epsilon 2\epsilon 4$ or the $\epsilon 4\epsilon 4$ genotype.

The distribution of the APOC3 genotypes did not differ between males and females (P = 0.489) (Table 4). In males, levels of blood lipid were similar between S1/S1 and S1/S2 genotypes, whereas in females, adolescents with the S1/S1 genotype had lower levels of TC and TC-HCLc compared with those with the S1/S2 genotype.

The distribution of the PPARγ2 genotypes did not differ between males and females (Pro12/Pro, 85 and 82% in males and females, respectively; Pro12/Ala, 15 and 18%, in males and females, respectively). No differences were found between the Pro12/Pro and Pro12/Ala12 genotypes in any of the lipid parameters studied, nor in BMI (data not shown). There were no differences between all the genotypes in birth weight in males and females. The outcome did not change when all the analyses were adjusted for age and/or pubertal development.

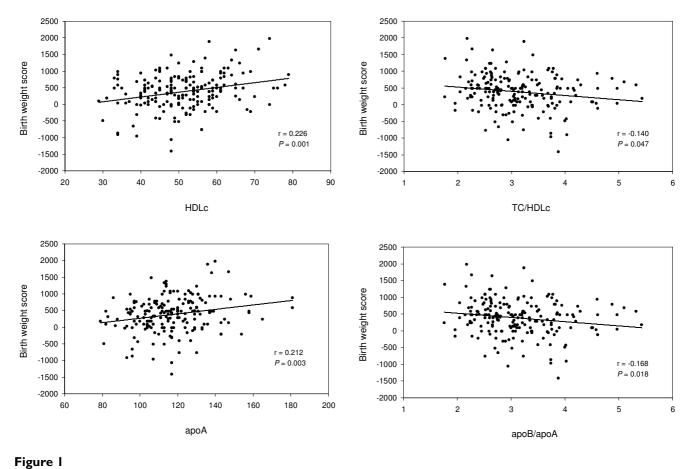
Birth weight, lipids and genotype interactions

Low birth weight was associated with higher levels of TC, LDLc, apoB, Lp(a), TC-HDLc, TC/HDLc and apoB/apoA in males with the APOE ϵ 3 ϵ 4 genotype, after controlling for age, pubertal status, socioeconomic status, gestational age, BMI and cardiorespiratory fitness (Table 5). Low birth weight was also associated with lower levels of HDLc and apoA in males with the APOE ϵ 3 ϵ 3 genotype. In females with the APOE ϵ 3 ϵ 4 genotype, low birth weight was associated with lower levels of TG. Low birth weight was not significantly associated with any of the lipid parameters studied in those adolescents with the APOE ϵ 2 ϵ 3 or in females with the ϵ 3 ϵ 3 genotype.

Low birth weight was associated with lower levels of apoA in males with the APOC3 S1/S1 genotype, and higher levels of Lp(a) in those with both the APOC3 S1/S1 and the S1/S2 genotype. In females, low birth weight was not significantly associated with blood lipids in any of the APOC3 genotypes (Table 6). There were no associations between low birth weight and blood lipids in any of the PPAR γ 2 genotypes (data not shown). The outcome did not change when BMI was not included in the model. The results did not materially change either when another anthropometric index (i.e. sum of six skinfold thicknesses, percentage of body fat, fat free mass, waist circumference, height squared) rather than BMI was included in the model.

Discussion

The results of the present study show that low birth weight seems to have a deleterious influence on blood lipid profile, particularly in those adolescents with the APOE $\varepsilon 3/\varepsilon 4$ genotype. The results also suggest that low birth weight is associated with lower levels of HDLc and apoA, and higher TC/HDLc and apoB/apoA ratios in male adolescents. Yet, low birth weight is not associated with blood lipid levels in female adolescents. Out of the three genes analyzed, the APOE genotypes seem to be a major determinant factor of lipid and lipoprotein profile in both sexes. Taken together, these results support the notion that intrauterine environment interact with genes in determining blood lipid profile in later life. One possible explanation to these findings is that a reduced foetal growth may have negative consequences on the liver growth. A poor liver growth may cause a down-regulation of the hepatic receptors, as well as disturbances in the synthesis of cholesterol. Consequently, the influence of the APOE genotypes on the lipid and lipoprotein metabolism



Birth weight score (g) and blood lipid and lipoprotein levels (mg/dl) in male adolescents. HDLc indicates high density lipoprotein cholesterol; apo, apolipoprotein; TC, total cholesterol.

would be intensified in those individuals with a low birth weight.

The potential underlying mechanism linking low birth weight with and adverse lipid profile in later life remains unclear. Barker et al. [7] suggested that raised serum TC, LDLc, and apoB levels in adult life might be associated with impaired growth during late gestation, when fetal undernutrition has a disproportionate effect on liver growth. Therefore, as the liver regulates lipid metabolism, impaired in utero growth of the liver may program a more adverse lipid profile.

Interpretation of the birth weight effect on blood lipids after adjustment for current body size is controversial, and available results indicate that postnatal changes in size, rather than fetal growth, is important [35]. Our findings show that the influence of low birth weight on HDLc, apoA, TC/HDLc and apoB/apoA in males was not altered after controlling for any of the measures of current body size or by others potential confounders such as age [15],

pubertal maturation [27], cardiorespiratory fitness [14,36] or socioeconomic status [37]. It is noteworthy that the associations between birth weight and blood lipids were mainly observed in males. A number of studies have found evidences for a sex related difference in the programming effect of low birth weight and later cardiovascular risk factors [3,38]. Sex related differences in the association between fetal growth and later coronary heart disease risk suggest that male fetuses are more vulnerable to the effects of fetal undernutrition [38]. One of the explanations lays on the fact that male fetuses seem to grow at a faster rate than female foetuses [39,40]. Therefore, assuming that fetal nutritional programming is the primary pathway for the association between birth weight and later cardiovascular disease risk factors it could be expected these associations to be stronger in males compared to females [39,40]. Our results concur with those showed in a meta-analyses by Lawlor et al. [38]. They reported that the association between birth weight and total cholesterol was stronger in males compared to females. The biological explanation for the different out-

Genotype n (%)		Males (n =	260)	Females (n = 242)				
	ε2/ε3 22 (8.5)	ε3/ε3 186 (71.6)	ε3/ε4 52 (19.9)	P value	ε2/ε3 21 (8.5)	ε3/ε3 Ι 75 (72.6)	ε3/ε4 46 (18.9)	P value
TC**	144.2* ± 34.4	157.9 ± 24.8	160.6 ± 26.8	<0.05	160.8 ± 24.7	69. ± 25.	177.0 ± 30.8	NS
HDLc**	53.0 ± 8.6	51.4 ± 9.6	49.8 ± 10.3	NS	65.5* ± 11.1	58.6 ± 11.0	58.5 ± 13.1	NS
LDLc**	77.2 [†] ± 28.6	92.7 ± 22.9	95.3 ± 23.8	<0.01	83.2 [†] ± 24.7	97.5 ± 21.9	103.4 ± 26.1	<0.01
TG**	69.7 ± 28.7	69.4 ± 28.1	77.5 ± 39.0	NS	60.4 ± 22.1	64.4 ± 26.1	68.0 ± 30.0	NS
apoA ^{∞∗}	115.9 ± 15.6	8. ± 7.3	114.5 ± 16.3	NS	4 .7†± 5.6	125.7 ± 17.9	128.9 ± 23.3	<0.01
apoB**	55.6† ± 15.8	68.0 ± 13.0	68.8 ± 16.4	<0.001	59.3†±11.7	70.1 ± 12.9	73.9 ± 16.1	<0.001
Lp(a)**	23.1 ± 33.4	31.0 ± 35.1	33.1 ± 41.9	NS	14.3 ± 15.3	33.0 ± 40.3	37.6 ± 38.5	NS
TC-HDLc	91.1* ± 31.6	106.6 ± 24.3	110.8 ± 27.7	<0.01	95.3* ± 25.5	110.4 ± 24.0	118.5 ± 30.5	<0.01
TC/HDLc	2.7* ± 0.7	3.2 ± 0.7	3.4 ± 0.9	<0.01	2.5* ± 0.7	3.0 ± 0.6	3.1 ± 0.8	<0.01
ароВ/ароА	0.5* ± 0.1	0.6 ± 0.1	0.6 ± 0.2	<0.01	0.4 [†] ± 0.1	0.6 ± 0.1	0.6 ± 0.2	<0.001
BMI	20.9 ± 3.9	22.2 ± 3.8	22.4 ± 4.0	NS	21.5 ± 3.9	21.7 ± 3.6	21.8 ± 3.4	NS

Table 3: Lipid and lipoprotein levels (mg/dl), and body mass index (kg/m²) according to the APOE genotypes and sex

Values are expressed as mean \pm standard deviation, otherwise stated.

P value from ANOVA, adjusted for mass significance.

*P < 0.05; \dagger P < 0.01 for ϵ 2/ ϵ 3 vs ϵ 3/ ϵ 3 (Tukey test).

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo, apolipoprotein; Lp(a), lipoprotein (a); BMI, body mass index; NS, not significant.

**Analyses were performed on log transformed data, but non-transformed data are presented in the table.

comes obtained in males and females warrant further investigation. It has been reported that the influence of sex on genes involved in the lipid and glucose metabolism could be attributed to hormonal differences [41,42]. However, we did not measure sex hormones, which hamper a further study of hormone-lipoprotein relationships in the studied population.

Variation in blood lipid levels seems to be partially determined by the APOE genotypes [10], which agree with our data. APOE genotype has also been shown to affect the association between birth weight and blood lipids in children [10]. Garces et al. reported that a greater effect of the APOE genotype on TC, LDLc and apoB levels was found in children with low birth weight [10]. In our study, low birth weight was associated with higher levels of TC and LDLc, apoB, and Lp(a) in males with the APOE $\varepsilon 3/\varepsilon 4$ genotype. Likewise, low birth weight was also associated with higher apoB/apoA ratio, which is a strong cardiovascular risk factor [43]. This observation suggests that the interaction of APOE genotype and birth weight might be an important determinant for future atherosclerosis.

Genotype		Males (n = 260)		Females (n = 242)				
	SI/SI	SI/S2	P value	SI/SI	SI/S2	P value		
n (%)	214 (82.1)	46 (17.0)		198 (81.8)	44 (18.2)			
TC*	156.2 ± 26.4	159.7 ± 27.0	NS	167.2 ± 25.2	181.9 ± 28.9	<0.001		
HDLc*	51.0 ± 9.4	51.1 ± 11.4	NS	59.0 ± 11.4	60.0 ± 12.0	NS		
LDLc*	91.1 ± 23.7	93.3 ± 25.2	NS	95.5 ± 22.4	106.3 ± 25.7	NS		
TG*	70.5 ± 31.7	76.1 ± 31.3	NS	63.5 ± 26.1	70.6 ± 27.8	NS		
apoA*	7.6 ± 7.2	4. ± 6.5	NS	127.0 ± 19.5	130.2 ± 18.1	NS		
apoB*	66.6 ± 14.4	68.4 ± 14.0	NS	68.8 ± 13.1	74.9 ± 16.1	NS		
Lp(a)*	31.2 ± 37.1	27.9 ± 32.4	NS	32.5 ± 39.4	31.8 ± 36.8	NS		
TC-HDLc	105.2 ± 25.7	108.6 ± 27.0	NS	108.2 ± 23.9	121.8 ± 31.4	<0.01		
TC/HDLc	3.2 ± 0.7	3.3 ± 0.8	NS	2.9 ± 0.6	3.2 ± 0.9	NS		
ароВ/ароА	0.6 ± 0.1	0.6 ± 0.2	NS	0.6 ± 0.1	0.6 ± 0.2	NS		
BMI	22.1 ± 3.9	22.2 ± 3.8	NS	21.5 ± 3.4	22.6 ± 4.0	NS		

Values are expressed as mean ± standard deviation, otherwise stated.

P value from ANOVA, adjusted for mass significance.

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo,

apolipoprotein; Lp(a), lipoprotein (a); NS, not significant

*Analyses were performed on log transformed data, but non-transformed data are presented in the table.

	Males (n = 260)						Females (n = 242)					
	3	2/ε3	3	ε3/ε3 ε3/ε4		ε2/ε3		ε3/ε3		ε 3 /ε 4		
	$\hat{oldsymbol{eta}}$	P value	\hat{eta}	P value	\hat{eta}	P value	\hat{eta}	P value	\hat{eta}	P value	\hat{eta}	P value
TC*	0.019	0.976	0.198	0.065	-0.401	0.042	0.787	0.353	0.157	0.197	-0.313	0.262
HDLc*	0.487	0.242	0.285	0.011	0.262	0.238	0.797	0.427	0.153	0.195	0.525	0.027
LDLc*	-0.109	0.787	0.121	0.260	-0.622	0.003	0.458	0.566	0.116	0.354	-0.302	0.264
TG*	-0.023	0.956	0.001	0.988	0.048	0.824	0.610	0.031	-0.153	0.171	-0.477	0.039
apoA*	0.424	0.326	0.236	0.022	0.328	0.125	0.698	0.314	0.150	0.900	-0.267	0.278
apoB*	-0.092	0.820	0.160	0.145	-0.554	0.008	0.428	0.540	-0.172	0.172	-0.076	0.780
Lp(a)*	-0.064	0.863	0.122	0.259	-0.625	0.002	-0.301	0.743	0.054	0.667	0.054	0.843
TC-HDLc	-0.019	0.663	0.127	0.227	-0.454	0.024	0.518	0.522	0.085	0.501	-0.060	0.816
TC/HDLc	-0.402	0.286	0.014	0.889	-0.406	0.028	0.106	0.900	-0.041	0.742	0.243	0.291
apoB/apoA	-0.333	0.387	-0.007	0.946	-0.518	0.011	0.142	0.850	0.047	0.699	0.130	0.588

Table 5: Regression coefficients (β) showing the association between birth weight score and lipid and lipoprotein levels by APOE genotypes and sex after controlling for age, pubertal stage, socioeconomic status, gestational age, body mass index and cardiorespiratory fitness

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo, apolipoprotein; Lp(a), lipoprotein a.

*Analyses were performed on log transformed data.

Female adolescents with the APOC3 S1/S2 genotype had elevated TC and non-HDLc levels, which is in agreement with another study [44]. The levels of triglyceride were similar between S1/S1 and S1/S2 genotypes in both males and females as reported in other population samples [45,46]. We also found that low birth weight was associated with lower levels of apoA in males with the APOC3 S1/S1 genotype, whereas low birth weight was associated with higher levels of Lp(a) in males with both S1/S1 and S1/S2 genotypes. This is the first study examining the influence of this polymorphism on the association between low birth weight and blood lipid levels in adolescents.

The PPAR γ 2 gene plays an important role in the regulation of glucose, lipid and energy metabolism [47], therefore it would be expected that the PPAR γ 2 genotypes could affect circulating lipid levels. The findings concerning the associations between PPAR γ 2 genotypes and lipid levels are contradictory [11,41,48-50]. We did not find differences on blood lipid and lipoprotein levels between genotypes, nor did on birth weight. Pfab et al. reported

Table 6: Regression coefficients (eta) showing the association between birth weight score and lipid and lipoprotein levels by APOC3
genotypes and sex after controlling for age, pubertal stage, socioeconomic status, gestational age, body mass index and
cardiorespiratory fitness

		Males (n = 260)	Females (n = 242)				
	SI/SI		SI/S2		SI/SI		S1/S2	
	\hat{eta}	P value	$\hat{oldsymbol{eta}}$	P value	$\hat{oldsymbol{eta}}$	P value	$\hat{oldsymbol{eta}}$	P value
TC*	0.050	0.609	-0.301	0.211	0.032	0.795	-0.309	0.535
HDLc*	0.184	0.053	-0.148	0.493	-0.063	0.590	0.363	0.346
LDLc*	-0.089	0.360	-0.208	0.387	-0.002	0.987	-0.305	0.528
TG*	0.048	0.616	-0.190	0.405	0.188	0.113	-0.588	0.227
ароА*	0.244	0.010	-0.146	0.522	-0.120	0.279	0.281	0.456
apoB*	-0.043	0.668	-0.267	0.268	0.022	0.859	-0.330	0.469
Lp(a)*	-0.294	0.003	-0.731	<0.001	0.021	0.865	0.020	0.867
TC-HDLc	0.022	0.871	-0.278	0.277	0.050	0.682	-0.409	0.394
TC/HDLc	-0.086	0.355	-0.087	0.722	0.097	0.407	-0.433	0.338
ароВ/ароА	-0.144	0.130	-0.166	0.504	0.108	0.351	-0.377	0.367

TC indicates total cholesterol; HDLc, high density liprotein cholesterol; LDLc, low density lipoprotein cholesterol; TG, triglycerides; Apo, apolipoprotein; Lp(a), lipoprotein a.

*Analyses were performed on log transformed data.

that neither the fetal nor the maternal PPAR γ 2 genotypes affected the birth weight of the 1950 newborn studied [50]. There is one study underlying the influence of PPAR γ 2 genotypes on the association between birth weight and lipid levels among elderly people [11]. It was reported that the Ala12 allele was associated with an increased TC, LDLc, and non-HDLc levels only among those who had birth weights lower than 3000 g. We did not find a significant effect of birth weight on lipid levels regarding the analyzed PPAR γ 2 genotypes, which agree with others [50].

Other factors such as current dietary intake may have affected the lipid levels, but such data were not available in the AVENA Study. There is controversy about the influence of age at the onset of menses on lipid and lipoprotein levels. Yet, no effect of age of menarche on lipid levels have been observed in the AVENA Study population [15]. The inclusion of age of menarche in the analysis rather than pubertal status did not alter the outcome. Birth weight might be a crude marker of the influence of maternal environment on fetal development and therefore underestimate the influence of such factors on offspring health outcomes. Future studies should directly measure potentially modifiable factors such as maternal nutrition, smoking, alcohol consumption or exercise, and relate these to offspring cardiovascular disease risk factors. It also remains possible that other birth measures including ponderal index, head circumference of waist circumference could be more important, but these measures were not available in the present study. The results should be interpreted with caution due to the small sample sizes.

Conclusion

The results of this study suggest that intrauterine environment interact with the genetic background affecting the lipid profile in later life. The findings indicate that low birth weight has a deleterious influence on lipid profile particularly in adolescents with the APOE $\varepsilon 3/\varepsilon 4$ genotype.

Competing interests

The authors declare that they have no competing interests. No benefits in any form have been received or will be received from a commercial party related directly or indirectly to the subject of this article.

Authors' contributions

JRR conceived the hypothesis, conducted the statistical analyses, and drafted the manuscript. IL, FOP and LAM critically revised the drafted manuscript. DGL, AM, MGF, JAM performed the genetic analysis. All authors contributed to the interpretation and discussion of the results, and critically revised the drafted manuscript.

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Acknowledgements

The AVENA study was supported by the Spanish Ministry of Health Instituto de Salud Carlos III (FIS PI021830), the Spanish Ministry of Health, FEDER-FSE funds FIS n° 00/0015, CSD grants 05/UPB32/0, 109/UPB31/03

and 13/UPB20/04, the Spanish Ministry of Education (AP-2004-2745; EX-2007-1124), scholarships from Panrico S.A., Madaus S.A. and Procter and Gamble S.A.

References

- Barker DJ, Eriksson JG, Forsen T, Osmond C: Fetal origins of adult disease: strength of effects and biological basis. Int J Epidemiol 2002, 31(6):1235-1239.
- Labayen I, Moreno LA, Ruiz JR, Gonzalez-Gross M, Warnberg J, Breidenassel C, Ortega FB, Marcos A, Bueno M: Small birth weight and later body composition and fat distribution in adolescents: the Avena study. Obesity (Silver Spring) 2008, 16(7):1680-1686.
- Labayen I, Moreno LA, Blay MG, Blay VA, Mesana MI, Gonzalez-Gross M, Bueno G, Sarria A, Bueno M: Early programming of body composition and fat distribution in adolescents. J Nutr 2006, 136(1):147-152.
- Ortega FB, Ruiz JR, Labayen I, Redondo C, Breidenassel C, Gomez S, Moreno LA, Castillo MJ: High fitness is associated with a healthier programming of body composition at adolescence. Am J Hum Biol 2008.
- Ortega FB, Labayen I, Ruiz JR, Martin-Matillas M, Vicente-Rodriguez G, Redondo C, Warnberg J, Gutierrez A, Sjostrom M, Castillo MJ, et al.: Are Muscular and Cardiovascular Fitness Partially Programmed at Birth? Role of Body Composition. J Pediatr 2008.
- Martyn CN, Gale CR, Jespersen S, Sherriff SB: Impaired fetal growth and atherosclerosis of carotid and peripheral arteries. Lancet 1998, 352:173-178.
- Barker DJ, Martyn CN, Osmond C, Hales CN, Fall CH: Growth in utero and serum cholesterol concentrations in adult life. *Bmj* 1993, 307(6918):1524-1527.
- Miura K, Nakagawa H, Tabata M, Morikawa Y, Nishijo M, Kagamimori S: Birth weight, childhood growth, and cardiovascular disease risk factors in Japanese aged 20 years. Am J Epidemiol 2001, 153(8):783-789.
- Owen CG, Whincup PH, Odoki K, Gilg JA, Cook DG: Birth weight and blood cholesterol level: a study in adolescents and systematic review. Pediatrics 2003, 111(5 Pt 1):1081-1089.
- Garces C, Benavente M, Ortega H, Rubio R, Lasuncion MA, Rodriguez Artalejo F, Fernandez Pardo J, de Oya M: Influence of birth weight on the apo E genetic determinants of plasma lipid levels in children. Pediatr Res 2002, 52(6):873-878.
- Eriksson J, Lindi V, Uusitupa M, Forsen T, Laakso M, Osmond C, Barker D: The effects of the Prol 2Ala polymorphism of the PPARgamma-2 gene on lipid metabolism interact with body size at birth. Clin Genet 2003, 64(4):366-370.
- Eriksson JG, Lindi V, Uusitupa M, Forsen TJ, Laakso M, Osmond C, Barker DJ: The effects of the Pro12Ala polymorphism of the peroxisome proliferator-activated receptor-gamma2 gene on insulin sensitivity and insulin metabolism interact with size at birth. Diabetes 2002, 51(7):2321-2324.
- Labayen I, Moreno LA, Marti A, Gonzalez-Lamuno D, Warnberg J, Ortega FB, Bueno G, Nova E, Ruiz JR, Garagorri JM, et al.: Effect of the Ala12 allele in the PPARgamma-2 gene on the relationship between birth weight and body composition in adolescents: the AVENA study. *Pediatr Res* 2007, 62(5):615-619.
- cents: the AVENA study. Pediatr Res 2007, 62(5):615-619.
 14. Castillo-Garzon M, Ruiz JR, Ortega FB, Gutierrez-Sainz A: A mediterranean diet is not enough for health: physical fitness is an important additional contributor to health for the adults of tomorrow. World Rev Nutr Diet 2007, 97:114-138.
- Ruiz JR, Ortega FB, Moreno LA, Wärnberg J, Gonzalez-Gross M, Cano MD, Gutierrez A, Castillo MJ, the AVENA Study Group: Serum lipid and lipoprotein reference values of Spanish adolescents; The AVENA study. Soz Praventivmed 2006, 51:99-109.
- McConathy WJ, Gesquiere JC, Bass H, Tartar A, Fruchart JC, Wang CS: Inhibition of lipoprotein lipase activity by synthetic peptides of apolipoprotein C-III. J Lipid Res 1992, 33(7):995-1003.
- 17. Semple RK, Chatterjee VK, O'Rahilly S: **PPAR gamma and human metabolic disease.** J Clin Invest 2006, **116(3):**581-589.
- Gonzalez-Gross M, Castillo MJ, Moreno L, Nova E, Gonzalez-Lamuno D, Perez-Llamas F, Gutierrez A, Garaulet M, Joyanes M, Leiva A, et al.: Alimentacion y valoracion del estado nutricional de los adolescentes espanoles (estudio AVENA). Evaluacion de riesgos y propuesta de intervencion. I. descripcion metodologica del proyecto. Nutricion Hospitalaria 2003, 18(1):15-28.

- Rodriguez G, Samper MP, Ventura P, Moreno LA, Olivares JL, Perez-Gonzalez JM: Gender differences in newborn subcutaneous fat distribution. Eur J Pediatr 2004, 163(8):457-461.
- Moreno LA, Mesana MI, Fleta J, Ruiz JR, Gonzalez-Gross M, Sarria A, Marcos A, Bueno M, Group AS: Overweight, obesity and body fat composition in spanish adolescents. The AVENA Study. Ann Nutr Metab 2005, 49(2):71-76.
- Moreno LA, Mesana MI, Gonzalez-Gross M, Gil CM, Fleta J, Warnberg J, Ruiz JR, Sarria A, Marcos A, Bueno M: Anthropometric body fat composition reference values in Spanish adolescents. The AVENA Study. Eur J Clin Nutr 2006, 60(2):191-196.
- Slaughter MH, Lohman TG, Boileau RA, Horswill CA, Stillman RJ, Van Loan MD, Bemben DA: Skinfold equations for estimation of body fatness in children and youth. Hum Biol 1988, 60(5):709-723.
- Tanner JM, Whitehouse RH: Clinical longitudinal standards for height, weight, height velocity, weight velocity, and stages of puberty. Arch Dis Child 1976, 51(3):170-179.
- Leger L, Lambert J, Goulet A, Rowan C, Dinelle Y: [Aerobic capacity of 6 to 17-year-old Quebecois – 20 meter shuttle run test with I minute stages]. Can | Appl Sport Sci 1984, 9(2):64-69.
- with I minute stages]. Can J Appl Sport Sci 1984, 9(2):64-69.
 25. Ortega FB, Ruiz JR, Castillo MJ, Moreno LA, Gonzalez-Gross M, Warnberg J, Gutierrez A: [Low level of physical fitness in Spanish adolescents. Relevance for future cardiovascular health (AVENA study)]. Rev Esp Cardiol 2005, 58(8):898-909.
- Kwiterovich PO Jr: Plasma lipid and lipoprotein levels in childhood. Ann N Y Acad Sci 1991, 623:90-107.
- Ruiz JR, Ortega FB, Tresaco D, Warnberg J, Mesa JL, Gonzalez-Gross M, Moreno LA, Marcos A, Gutierrez A, Castillo MJ: Serum lipids, body mass index and waist circumference during pubertal development in Spanish adolescents: the AVENA Study. Horm Metab Res 2006, 38(12):832-837.
- Higuchi R: Simple and rapid preparation of samples for PCR. In PCR Technology Edited by: Erlich H. Stockton Press; 1989:31-37.
- Hixson JE, Vernier DT: Restriction isotyping of human apolipoprotein E by gene amplification and cleavage with Hhal. J Lipid Res 1990, 31(3):545-548.
- Hixson JE, Henkel RD, Britten ML, Vernier DT, deLemos RA, Berg JL Vande, Walsh RA: Alpha-myosin heavy chain cDNA structure and gene expression in adult, fetal, and premature baboon myocardium. J Mol Cell Cardiol 1989, 21(10):1073-1086.
- Goyenechea E, Dolores Parra M, Alfredo Martinez J: Weight regain after slimming induced by an energy-restricted diet depends on interleukin-6 and peroxisome-proliferator-activatedreceptor-gamma2 gene polymorphisms. Br J Nutr 2006, 96(5):965-972.
- Dubois M, Pattou F, Kerr-Conte J, Gmyr V, Vandewalle B, Desreumaux P, Auwerx J, Schoonjans K, Lefebvre J: Expression of peroxisome proliferator-activated receptor gamma (PPARgamma) in normal human pancreatic islet cells. Diabetologia 2000, 43(9):1165-1169.
- Holm S: A simple sequentially rejective multiple test procedure. Scand J Statist 1979, 6:65-70.
- 34. Gordi T, Khamis H: Simple solution to a common statistical problem: interpreting multiple tests. *Clin Ther* 2004, **26(5)**:780-786.
- 35. Lucas A, Fewtrell MS, Cole TJ: Fetal origins of adult disease-the hypothesis revisited. *Bmj* 1999, 319(7204):245-249.
- Mesa JL, Ruiz JR, Ortega FB, Warnberg J, Gonzalez-Lamuno D, Moreno LA, Gutierrez A, Castillo MJ: Aerobic physical fitness in relation to blood lipids and fasting glycaemia in adolescents: Influence of weight status. Nutr Metab Cardiovasc Dis 2006, 16(4):285-293.
- Manhem K, Dotevall A, Wilhelmsen L, Rosengren A: Social gradients in cardiovascular risk factors and symptoms of Swedish men and women: the Goteborg MONICA Study 1995. J Cardiovasc Risk 2000, 7(5):359-368.
- Lawlor DA, Owen CG, Davies AA, Whincup PH, Ebrahim S, Cook DG, Davey Smith G: Sex differences in the association between birth weight and total cholesterol. A meta-analysis. Ann Epidemiol 2006, 16(1):19-25.
- Godfrey KM: Maternal regulation of fetal development and health in adult life. Eur J Obstet Gynecol Reprod Biol 1998, 78(2):141-150.
- 40. Lampl M, Jeanty P: Timing is everything: a reconsideration of fetal growth velocity patterns identifies the importance of

individual and sex differences. Am J Hum Biol 2003, 15(5):667-680.

- Dedoussis GV, Theodoraki EV, Manios Y, Yiannakouris N, Panagiotakos D, Papoutsakis C, Skenderi K, Zampelas A: The Pro12Ala polymorphism in PPARgamma2 gene affects lipid parameters in Greek primary school children: A case of gene-to-gender interaction. Am J Med Sci 2007, 333(1):10-15.
- Thamer C, Haap M, Volk A, Maerker E, Becker R, Bachmann O, Machicao F, Haring HU, Stumvoll M: Evidence for greater oxidative substrate flexibility in male carriers of the Pro 12 Ala polymorphism in PPARgamma2. Horm Metab Res 2002, 34(3):132-136.
- Yusuf S, Hawken S, Ounpuu S, Dans T, Avezum A, Lanas F, McQueen M, Budaj A, Pais P, Varigos J, et al.: Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. Lancet 2004, 364(9438):937-952.
- Dallongeville J, Meirhaeghe A, Cottel D, Fruchart JC, Amouyel P, Helbecque N: Gender related association between genetic variations of APOC-III gene and lipid and lipoprotein variables in northern France. Atherosclerosis 2000, 150(1):149-157.
- Dallongeville J, Cottel D, Montaye M, Codron V, Amouyel P, Helbecque N: Impact of APOA5/A4/C3 genetic polymorphisms on lipid variables and cardiovascular disease risk in French men. Int J Cardiol 2006, 106(2):152-156.
- 46. Russo GT, Meigs JB, Cupples LÁ, Demissie S, Otvos JD, Wilson PW, Lahoz C, Cucinotta D, Couture P, Mallory T, et al.: Association of the Sst-I polymorphism at the APOC3 gene locus with variations in lipid levels, lipoprotein subclass profiles and coronary heart disease risk: the Framingham offspring study. Atherosclerosis 2001, 158(1):173-181.
- Stumvoll M, Haring H: The peroxisome proliferator-activated receptor-gamma2 Prol2Ala polymorphism. Diabetes 2002, 51(8):2341-2347.
- Deeb SS, Fajas L, Nemoto M, Pihlajamaki J, Mykkanen L, Kuusisto J, Laakso M, Fujimoto W, Auwerx J: A Prol 2Ala substitution in PPARgamma2 associated with decreased receptor activity, lower body mass index and improved insulin sensitivity. Nat Genet 1998, 20(3):284-287.
- Meirhaeghe A, Fajas L, Helbecque N, Cottel D, Auwerx J, Deeb SS, Amouyel P: Impact of the Peroxisome Proliferator Activated Receptor gamma2 Pro I 2Ala polymorphism on adiposity, lipids and non-insulin-dependent diabetes mellitus. Int J Obes Relat Metab Disord 2000, 24(2):195-199.
- Pfab T, Poralla C, Richter CM, Godes M, Slowinski T, Priem F, Halle H, Hocher B: Fetal and maternal peroxisome proliferatoractivated receptor gamma2 Prol2Ala does not influence birth weight. Obesity (Silver Spring) 2006, 14(11):1880-1885.

Pre-publication history

The pre-publication history for this paper can be accessed here:

http://www.biomedcentral.com/1471-2350/9/98/prepub

