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3 **TITLE**

4
5 **Antimicrobial susceptibility of *Helicobacter pylori* against six currently**
6 **used antibiotics in Spain**

7 Running title: Searching for appropriate *H. pylori* eradication antibiotics

8
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52 **SYNOPSIS**

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54 **Background:** Antibiotic resistance is directly related to the loss of efficacy of
55 currently accepted *Helicobacter pylori* therapies. The knowledge of the antibiotic
56 susceptibility in a local area can contribute to design specific "à la carte" treatments.
57 The aim of this study was to analyze the susceptibility pattern of *H. pylori* isolates
58 regarding to six conventional antibiotics currently used in a northern region of
59 Spain.

60 **Materials and methods:** Seventy-one isolates were obtained from gastric biopsies
61 of 76 consecutive adult patients suffering from peptic ulcer disease, dyspepsia or
62 familiar gastric cancer and known to be infected with *H. pylori* by conventional
63 methods. Susceptibility testing was performed for amoxicillin, ciprofloxacin,
64 levofloxacin, clarithromycin, metronidazole, and tetracycline by using the Etest
65 method.

66 **Results:** The prevalence rates of resistance were as follows: amoxicillin, 1.4%
67 (95% Confidence Interval [CI], 0.0 to 7.6); clarithromycin, 14.7% (CI, 7.3 to 25.4);
68 ciprofloxacin, 14.3% (CI, 7.1 to 24.7); levofloxacin, 14.5% (CI, 7.2 to 25.0);
69 metronidazole, 45.1% (CI, 33.2 to 57.3); and tetracycline, 0% (CI, 0.0 to 5.1).

70 **Conclusions:** Our study confirms an increasing rate of resistance to levofloxacin
71 which equals that of clarithromycin in our health care area. This fact may reflect a
72 wide and indiscriminate use of the former antibiotic and could account for a loss of
73 clinical effectiveness of levofloxacin-containing regimens. Moreover, resistance rates
74 against clarithromycin remain stable which could allow us to maintain its use in our
75 area.

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78 INTRODUCTION

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80 *Helicobacter pylori* infects the gastric mucosa and represents the main cause of
81 gastritis, peptic ulcer disease, and gastric cancer. It has been demonstrated that
82 eradication of *H. pylori* improves the clinical outcome of patients with duodenal
83 ulcer, prevents its recurrence, and decrease the risk of gastric cancer in infected
84 patients.^{1,2} Nowadays, the European and American guidelines on the treatment of
85 *H. Pylori* infection recommend as first-line therapy a combination of a proton pump
86 inhibitor (PPI) with two antibiotics, being omeprazole and clarithromycin plus
87 amoxicillin or metronidazole the preferred regimen.^{3,4} Although the initial eradication
88 success rate for the standard triple therapy was in excess of 90% ten years ago,
89 therapy failures up to 30-40% of cases using this regimen have been more recently
90 reported.⁵⁻⁸ Even though patient's lack of compliance, inadequate length of therapy,
91 or a high bacterial burden are conditions that may contribute to such loss of
92 efficacy, antimicrobial resistance is regarded as the leading factor responsible for
93 eradication failure. This issue is of particular relevance in regard to clarithromycin
94 which can induce a virtually 70% loss of effectiveness when takes part of an PPI-
95 amoxicillin based triple therapy, depending on *in vitro* macrolide susceptibility.⁹ In
96 fact, the most recent Maastricht guidelines on *H. pylori* infection management
97 recommend substituting metronidazole for clarithromycin when resistance to this
98 antibiotic exceeds 15-20%.³ Finally, there is a wide geographic variation regarding
99 the prevalence of antibiotic resistance. This fact has been recently highlighted in the
100 updated European Surveillance of *H. pylori* Resistance to Antibiotics where
101 differences in clarithromycin resistance rates of more than 10% were detected
102 between different regions of Europe, precluding its use in some of them.¹⁰ All of
103 that makes general guidelines related to the use of different antibiotic regimens

104 against *H. pylori* useless if they do not include the available data about antibiotic
105 susceptibility in local areas. Taking into account the aforementioned considerations
106 it seems desirable to have regularly updated, reliable information on the prevalence
107 of antibiotic resistance to *H. pylori* for the various countries, regions or health care
108 areas. Such information can aid to establish on an individual basis the potentially
109 most effective eradicating regimen for *H. pylori* infection.¹¹⁻¹³
110 The aim of this study was to assess the susceptibility of *H. pylori* strains isolated
111 from gastric biopsies of patients with gastroduodenal peptic ulcer disease, treatment
112 unresponsive dyspepsia, or family history of gastric cancer to six antibiotics
113 commonly used in therapeutic procedures.

114 MATERIAL AND METHODS

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116 Patients and sample collection

117 From February to December, 2010, seventy-six consecutive adult patients who had
118 not been previously eradicated against *H. pylori* were evaluated at the
119 Gastroenterology Department of the Hospital of Laredo, a community hospital
120 placed on the North of Spain. These patients referred different upper abdominal
121 complaints, a familiar history of gastric cancer and/or a personal history of
122 gastroduodenal peptic ulcer disease. All of them underwent a diagnostic
123 esophagogastroduodenoscopy including collection of biopsies of the gastric mucosa
124 (from both the body and the antrum) for rapid urease test, histologic study, and
125 culture.

126

127 Culture preparation and susceptibility testing

128 The biopsy specimens of those patients with a positive urease test were
129 homogenized and sowed in selective (Agar Pylori-BioMérieux, Sweden) and no
130 selective (Columbia III Agar with 5% Sheep Blood, Becton-Dickinson, Germany)
131 culture media and incubated at 37°C under microaerophilic conditions for 10-14
132 days. Once cultures were obtained, the isolates were identified according to colonial
133 morphology, Gram-staining, urease, catalase and oxidase tests. Afterwards, the
134 strains were re-sowed and subcultured in non-selective media (Columbia III Agar
135 with 5% Sheep Blood) for 48-72 hours in order to perform the susceptibility studies.
136 The minimum inhibitory concentration (MIC) was determined using the Etest
137 method as recommended by the *British Society for Antimicrobial Chemotherapy*
138 (BSAC).¹⁴ Muller Hinton agar supplemented with 5% sheep blood (Becton-Dickinson,
139 Germany) was used as culture media. The *H. pylori* culture suspension of 3.0

McFarland turbidity was used to inoculate the plates by confluent swabbing and Etest strips (BioMérieux, Sweden) were applied onto culture plates. The plates were incubated at 35°C for 3-5 days under microaerophilic conditions. The tested drugs were amoxicillin, clarithromycin, ciprofloxacin, levofloxacin, metronidazole, and tetracycline. The breakpoints used to classify strains as susceptible or resistant according to the MIC value were as follows: ≤ 1 mg/L, susceptible (S) and ≥ 2 mg/L, resistant (R) for amoxicillin and clarithromycin; ≤ 1 mg/L, S and ≥ 1 mg/L, R for ciprofloxacin and levofloxacin; ≤ 4 mg/L, S and ≥ 8 mg/L, R for metronidazole; and ≤ 2 mg/L, S and ≥ 4 mg/L, R for tetracycline. The breakpoints for amoxicillin, clarithromycin, metronidazole, and tetracycline were interpreted according to the BSAC recommendations. Quinolones are not standardised by the BSAC so we used those recommended by the *Société Française de Microbiologie* that are in accordance with those suggested by other authors.^{15,16}

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154 **Statistics**

155 A descriptive analysis was performed using the v.15.0 SPSS package.

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157 **Ethics**

158 This study was performed following the current standards of Good Clinical Practice
159 and Good Laboratory Practice and the protocol was approved by the Cantabric
160 Ethical Investigation Committee. An informed consent was obtained from all the
161 patients.

RESULTS

Seventy-one *H. pylori* strains were isolated from 76 consecutive adult patients (31 male and 45 female) who had been included in the study on the basis of a positive rapid urease test. Susceptibility figures corresponding to the six tested antimicrobials were determined according to the aforementioned Etest method and are summarised in Table 1 (in 4 of 71 isolates antimicrobial susceptibility could not be tested for all antibiotics).

Forty-four out of 71 isolates (62%) showed resistance to at least one antibiotic, while in 27 patients all isolates were susceptible to the tested antibiotics. Resistance to only one antibiotic was present in the isolates obtained from thirty-one patients. On the other hand, 13 isolates showed resistance to 2 or more antibiotics, mainly involving quinolones (5 patients). Isolates coming from five patients showed multi-resistance, defined as having resistance to 3 or more antibiotics (in four patients, *H. pylori* strains exhibited resistance to ciprofloxacin, levofloxacin and metronidazole, and in another patient the isolate was also resistant to clarithromycin). Finally, only 1 patient had an isolate that was simultaneously resistant to levofloxacin and clarithromycin.

183 **DISCUSSION**

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185 The knowledge of the available data about *in vitro* antimicrobial susceptibility of *H.*
186 *pylori* has been requested by several experts in order to increase the therapeutic
187 success rate for *H. pylori* eradication.^{11,13} The rationale for this claim relies on two
188 facts: first, the well-known variability on the prevalence of antibiotic resistance
189 among different countries or even regions or social groups (mainly related to
190 clarithromycin); and second, the special consequences that a wrong antibiotic
191 selection can bear, as it has been stated before (for example, a nearly 70% loss of
192 efficacy depending on susceptibility or resistance to clarithromycin).

193 In the present study, we have found a wide spectrum of resistance rates of *H.*
194 *pylori*, from nearly negligible figures against tetracycline (0%) and amoxicillin
195 (1.4%) to high resistance rates against metronidazole (45.1%). Intermediate and
196 virtually identical figures were found for clarithromycin (14.7%), ciprofloxacin
197 (14.3%), and levofloxacin (14.5%). These results merit some considerations. Firstly,
198 the prevalence rates of *H. pylori* resistance against clarithromycin and amoxicillin
199 are in accordance to those reported in 2001 in Madrid (Spain) within the European
200 Multicentre Survey of *in vitro* Antimicrobial Resistance in *H. pylori* (15% and 0%,
201 respectively).¹⁷ It is important to remark the scarce variation on resistance patterns
202 found throughout the last decade, particularly in regard to clarithromycin, which
203 could stand for a maintained relatively high efficacy of the standard triple therapy
204 against *H. pylori* in our region. These results are in accordance with those reported
205 in a recent European survey, where clarithromycin resistance rates were 17% as a
206 whole.¹⁰ While countries from de Centre, West, and South of Europe have
207 experienced a great increase in resistance rates against clarithromycin (>20%),
208 which jeopardizes its use as part of conventional triple therapy empirical regimens,

northern countries and other exceptions like Germany and Spain maintain low-intermediate resistance rates. Our group is **in the process of** conducting a randomised clinical trial on first-line *H. pylori* eradication therapy comparing two triple therapy based regimens (namely omeprazole, amoxicillin, and either clarithromycin or levofloxacin) which will help us to clarify the true clinical effectiveness of clarithromycin and levofloxacin in our area. A high rate of eradication with standard first-line therapies has been achieved in other regions on the basis of a high adherence to the treatment so a combination of moderate slowly-growing rate of resistance to antibiotics and high levels of compliance could account for a sustained high eradication rates.^{8,18} **Secondly**, the resistance rates to metronidazole are slightly higher than previously reported in Spain (37.2%) and in Europe (33.1%) in 2001, approaching to those encountered in Italy (49%), Austria (44.9%), and Greece (44.1%).¹⁷ Certainly, metronidazole resistances have remained stable in the last decade in Europe as a whole (34.3%; CI, 16.7 to 50.3)¹⁰ although individual figures of each country are awaited and differences between them as those found for clarithromycin can be expected to confirm our results. Even though *in vitro* resistance to metronidazole may not accurately reflect *in vivo* resistance, regimens including metronidazole could not be a preferable choice in populations with >40% metronidazole resistance.^{12,19} Thus, our data could dissuade gastroenterologists of our **region** to use this antibiotic as taking part of an alternative first-line therapeutic regimen against *H. pylori*, particularly in cases of penicillin allergy.

Thirdly and of great interest in this study is the notable *H. pylori* resistance rate against levofloxacin, a quinolone increasingly used as a clarithromycin-substitute for either first-line or rescue therapy in different regimens.²⁰ In spite of the high eradication rates (~90%) achieved with the combination of PPI, amoxicillin, and

235 levofloxacin, there are concerns about an increasing rate of quinolone resistance:
236 15% in Japan, 16.8% in Belgium, 23.1% in Italy, from 2.8% to 11.8% between
237 1998 and 2003 in Taiwan, from 3% in 1999 to 15% in 2004 in France, and from
238 11.2% in 2003 to 22.1% in 2005 in Germany.^{8,16,20,21-24} These changes on *H. pylori*
239 susceptibility to quinolones could account for a fall in the success rate of a triple
240 therapy including levofloxacin. In fact, some investigators have linked the slight
241 reduction in overall eradication rates of a levofloxacin-based re-treatment (from 76-
242 85.7% to 72.7%) to the high prevalence of *in vitro* primary resistance (30.3%)
243 which doubled that found in previous trials.²⁰ Although a relatively low rate of *H.*
244 *pylori* resistance to quinolones (6%) had been previously reported in Spain,²⁵ the
245 present study reveals and confirms an increasing rate of levofloxacin resistance in
246 our country which is similar to that has been reported in other Mediterranean areas.
247 In this way, the aforementioned European study on antibiotic resistance of *H.*
248 *pylori*¹⁰ underscores a progressive trend to a higher resistance rates to levofloxacin
249 (similar to those encountered in our study) which can discourage the future use of
250 eradication regimens including this quinolone. Finally, resistances against amoxicillin
251 and tetracycline remain negligible.

252 To sum up, the present work shows stable *in vitro* resistance rates of *H. pylori* to
253 clarithromycin which could support its use as a part of *H. pylori* eradication
254 regimens in our area. In addition, it also confirms a quick increase of the *in vitro*
255 resistance rate to levofloxacin in our region which may discourage its use in
256 eradication regimens, at least as first-line treatment. Taking into account the
257 important variability of prevalence rates of *H. pylori* resistance to different
258 antibiotics (in time and space) and the consequences that this fact can bring on
259 therapy success we encourage regional Gastroenterology and Microbiology Societies

260 to periodically update their data on *in vitro* resistances and then make suitable
261 recommendations about the best desirable therapy.

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272 **TRANSPARENCY DECLARATION**

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274 **None to declare.**

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REFERENCES

1. Sugiyama T, Sakaki N, Kozawa H, et al. Sensitivity of biopsy site in evaluating regression of gastric atrophy after *Helicobacter pylori* eradication treatment. *Aliment Pharmacol Ther* 2002; **16** (Suppl 2): S187-90.
2. McColl KE. Clinical practice. *Helicobacter pylori* infection. *N Engl J Med* 2010; **362**: 1597-604.
3. Malfertheiner P, Megraud F, O'Morain C, et al. Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. *Gut* 2007; **56**: 772-81.
4. Chey WD, Wong BC. Practice Parameters Committee of the American College of Gastroenterology. American College of Gastroenterology guidelines on the management of *Helicobacter pylori* infection. *Am J Gastroenterol* 2007; **102**: 1808-25.
5. Kearney DJ, Brousal A. Treatment of *Helicobacter pylori* infection in clinical practice in the United States: results from 224 patients. *Dig Dis Sci* 2000; **45**: 265-71.
6. Kadayifci A, Buyukhatipoglu H, Cemil Savas M, et al. Eradication of *Helicobacter pylori* with triple therapy: an epidemiologic analysis of trends in Turkey over 10 years. *Clin Ther* 2006; **28**: 1960-6.
7. Gisbert JP. "Rescue" regimens after *Helicobacter pylori* treatment failure. *World J Gastroenterol* 2008; **14**: 5385-402.
8. O'Connor A, Gisbert J, O'Morain C. Treatment of *Helicobacter pylori* infection. *Helicobacter* 2009; **14** Suppl 1: 46-51.
9. Mégraud F. H pylori antibiotic resistance: prevalence, importance, and advances in testing. *Gut* 2004; **53**: 1374-84.

- 311 10. Mégraud F, Kist M, López-Brea M, et al. Surveillance of *Helicobacter pylori*
312 Resistance to Antibiotics in Europe 2008-2009. *Gastroenterology* 2011; **140**
313 (suppl.1): 1715.
- 314 11. Castro-Fernández M, Vargas-Romero J. Infection with *Helicobacter pylori*.
315 Prevalence, research and impact of antibiotic resistance. *Rev Esp Enferm Dig* 2009;
316 **101**: 743-56.
- 317 12. Katelaris PH. *Helicobacter pylori*: antibiotic resistance and treatment options. *J*
318 *Gastroenterol Hepatol* 2009; **24**: 1155-7.
- 319 13. Ahmad N, Zakaria WR, Mohamed R. Analysis of antibiotic susceptibility patterns
320 of *Helicobacter pylori* isolates from Malaysia. *Helicobacter* 2011; **16**: 47-51.
- 321 14. King A. Recommendations for susceptibility tests on fastidious organisms and
322 those requiring special handling. *J Antimicrob Chemother* 2001; **48** Suppl 1: 77-80.
- 323 15. Comité de l'Antibiogramme de la Société Française de Microbiologie:
324 Recommandations du CASFM, Communiqué 2008 (Edition de Janvier 2008).
325 http://www.sfm-microbiologie.org/UserFiles/file/CASFM/casfm_2008.pdf
326 (accessed on May 20, 2011)
- 327 16. Bogaerts P, Berhin C, Nizet H, et al. Prevalence and mechanisms of resistance
328 to fluoroquinolones in *Helicobacter pylori* strains from patients living in Belgium.
329 *Helicobacter* 2006; **11**: 441-5.
- 330 17. Glupczynski Y, Mégraud F, López-Brea M, et al. European multicentre survey of
331 in vitro antimicrobial resistance in *Helicobacter pylori*. *Eur J Clin Microbiol Infect Dis*
332 2001; **20**: 820-3.
- 333 18. Seppälä K, Kosunen TU, Veijola L, et al. Cure of *Helicobacter pylori* infection in
334 all compliant patients: report on 644 subjects. *Scand J Gastroenterol* 2008; **43**:
335 1149-50.

- 336 19. Mégraud F. Epidemiology and mechanism of antibiotic resistance in
337 *Helicobacter pylori*. *Gastroenterology* 1998; **115**: 1278-82.
- 338 20. Perna F, Zullo A, Ricci C, et al. Levofloxacin-based triple therapy for
339 *Helicobacter pylori* re-treatment: role of bacterial resistance. *Dig Liver Dis* 2007; **39**:
340 1001-5.
- 341 21. Tankovic J, Lascols C, Sculo Q, et al. Single and double mutations in *gyrA* but
342 not in *gyrB* are associated with low- and high-level fluoroquinolone resistance in
343 *Helicobacter pylori*. *Antimicrob Agents Chemother* 2003; **47**: 3942-4.
- 344 22. Miyachi H, Miki I, Aoyama N, et al. Primary levofloxacin resistance and *gyrA/B*
345 mutations among *Helicobacter pylori* in Japan. *Helicobacter* 2006; **11**: 243-9.
- 346 23. Cattoir V, Nectoux J, Lascols C, et al. Update on fluoroquinolone resistance in
347 *Helicobacter pylori*: new mutations leading to resistance and first description of a
348 *gyrA* polymorphism associated with hypersusceptibility. *Int J Antimicrob Agents*
349 2007; **29**: 389-96.
- 350 24. Glocker E, Stueger HP, Kist M. Quinolone resistance in *Helicobacter pylori*
351 isolates in Germany. *Antimicrob Agents Chemother* 2007; **51**: 346-9.
- 352 25. Cibrelus LA, Pérez de Ayala A, Alarcón T, et al. In vitro efficiency of
353 ciprofloxacin and rifampicin as potential second-line treatment in Spanish
354 *Helicobacter pylori* clinical isolates. *Helicobacter* 2006; **11**: 402-3.
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362 **Table 1.** Antimicrobial susceptibility of *H. pylori* isolates in the North of Spain

Antibiotic	Isolates tested (n)	Isolates with resistance (n)	Resistance, % (95 CI)	MIC	
				S	R
Amoxicillin	71	1	1.4 (0.0-7.6)	≤1	≥2
Ciprofloxacin	70	10	14.3 (7.1-24.7)	≤1	≥1
Levofloxacin	69	10	14.5 (7.2-25)	≤1	≥1
Clarithromycin	68	10	14.7 (7.3-25.4)	≤1	≥2
Metronidazole	71	32	45.1 (33.2-57.3)	≤4	≥8
Tetracycline	71	0	0.0 (0.0-5.1)	≤2	≥4

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364 **MIC** = Minimal Inhibitory Concentration (mg/L). This parameter was established for each
365 antibiotic as breakpoints of susceptibility (S) or resistance (R); **n** = number; **95 CI** = 95%
366 Confidence Interval.

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