



ORIGINAL ARTICLE

Effects of vaccination with *Brucella melitensis*, strains Rev 1 Δ eryCD and Rev 1, on the reproductive system of young male goats

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Abstract The present study evaluates the effects of vaccination with *Brucella melitensis* strains Rev 1 Δ eryCD and Rev 1 on the reproductive system of male goats. Three groups, each of them consisting of 15 six-month-old brucellosis-free male goats, were studied. The first group was vaccinated with the Rev 1 Δ eryCD strain, the second group received Rev 1 and the third group was inoculated with sterile physiological saline solution. The dose of both strains was of 1×10^9 CFU/ml. Over the course of the five months of this study, three males from each group were euthanized every month. Their reproductive tracts, spleens, and lymph nodes were collected to analyze serology, bacteriology PCR, histology, and immunohistochemistry. Results show that vaccination with *B. melitensis* strains Rev 1 Δ eryCD and Rev 1 does not harm the reproductive system of male goats. Strain *B. melitensis* Rev 1 Δ eryCD displayed a lower capacity to colonize the reproductive tract than strain Rev 1, which was attributed to its limited catabolic action toward erythritol.

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PALABRAS CLAVE

Brucelosis;
Machos cabríos;
Vacunación;
Rev 1;
Rev 1 Δ eryCD

Efectos de la vacunación con *Brucella melitensis* cepas Rev 1 Δ eryCD y Rev 1 en el sistema reproductivo de caprinos machos jóvenes

Resumen La presente investigación evalúa los efectos de la vacunación con *Brucella melitensis* cepas Rev 1 Δ eryCD y Rev 1 en el sistema reproductivo de machos caprinos. Se estudiaron tres grupos de caprinos, integrado cada grupo por 15 machos libres de brucelosis de seis meses de edad. El primer grupo fue vacunado con la cepa Rev 1 Δ eryCD, el segundo grupo recibió Rev 1 y el tercer grupo recibió solución salina fisiológica estéril (control). El título de las dos cepas inoculadas fue de 1×10^9 UFC/ml. Tres machos de cada grupo fueron sacrificados cada mes durante los cinco meses de este estudio. Se recolectaron el tracto reproductivo, el bazo y los ganglios linfáticos para análisis de serología, bacteriología, PCR, histología e inmunohistoquímica. Los resultados muestran que la vacunación con las cepas Rev 1 Δ eryCD y Rev 1 de *B. melitensis* no causa daño al sistema reproductivo de los machos cabríos. La cepa *B. melitensis* Rev 1 Δ eryCD mostró menos capacidad para colonizar el tracto reproductivo que la cepa Rev 1 debido a su acción catabólica limitada hacia el eritritol.

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Introduction

Goats were one of the first domesticated animals, around 10 500 years ago in the Fertile Crescent. According to the FAO, the world goat population has been estimated to be around 921 million animals, with an increase of more than 20% in the last decade. Goats are a source of milk, meat, and fiber and are adapted to a wide range of grazing environments³¹. Approximately 90% of goats are found in the developing world, where they are considered one of the most important sources of protein for humans²³.

Brucella melitensis is the main causative agent of caprine brucellosis, which leads to significant production losses due to kid mortality and low milk production, and is also the main bacterial zoonosis in Mexico with a significant economic impact on public health²⁰.

Caprine brucellosis has been controlled in most industrialized countries; however, this disease remains endemic in resource-limited settings, where small ruminants are the major livestock species and the main economic livelihood, such as the Mediterranean region, the Middle East, Central Asia, sub-Saharan Africa, and parts of Latin America²³.

The Official Mexican Standard for the control of brucellosis specifies that goats should not be immunized with the *B. melitensis* Rev 1 vaccine¹⁷, as erythritol is known to promote the growth of *B. melitensis* Rev 1 and this strain has tropism for the male reproductive organs due to its content of erythritol^{7,15}. However, there is no scientific evidence to support that the Rev 1 vaccine causes lesions when applied to kids. From an epidemiological point of view, the use of vaccination against brucellosis in male goats would not be indicated in cases where the herd has low incidence or is free of the disease. Brucellosis in male goats occasionally causes epididymo-orchitis²⁴. Nevertheless, vaccination of kids could be an option in endemic areas of the disease to protect male goats from contracting brucellosis through frequent routes of infection such as licking the genitals of females, or consuming water or food contaminated with the

bacteria. In these cases, vaccination would protect against the eventual sacrifice of the kid due to the disease. So our working hypothesis was that the Rev 1 vaccine does not cause lesions on the male genitalia.

The *B. melitensis* Rev 1 Δ eryCD strain was constructed with the same characteristics as strain *B. abortus* S19. It presents a 702 bp deletion in specific sections of the *eryC* and *eryD* genes, inhibiting the ability of microorganisms to catabolize erythritol. Since the wild parental strain Rev 1 contains the complete *ery* operon, it is capable of metabolizing erythritol, which should contribute to the capacity of the strain to colonize the sexual organs. Hence, vaccination with the *B. melitensis* Rev 1 Δ eryCD strain, with the deleted *ery* operon, should show different effects on the male goat reproductive tract compared to those of vaccination with the wild parental Rev 1 strain. Therefore the present study aimed to compare the effects caused by vaccination with *B. melitensis* strains Rev 1 Δ eryCD and Rev 1 on the reproductive tract of young male goats.

Materials and methods**Vaccination**

Forty-eight four-month-old male goats of the Saanen breed were included in the study and were selected from brucellosis-free animals and tested negative in the serological study with the card test. To prevent the possible interference of the vaccine strains in the experiment, the three groups were separated after vaccination. The negative control group and the group vaccinated with Rev 1 were housed in separate pens with no possibility of contact in the facilities of the National Institute of Forestry, Agriculture and Livestock Research (INIFAP) in Mexico City. Concurrently, the kids of the group vaccinated with the Rev 1 Δ eryCD strain were housed in a ranch of the Faculty of Veterinary Medicine and Zootechnics of the National Autonomous University of Mexico, located in Chalco, State of Mexico. At

the beginning of the experiment, three unvaccinated kids were sacrificed to confirm through a serological and bacteriological study that they were negative for brucellosis. Three groups were formed, each one consisting of 15 male goats. The two treatment groups were vaccinated when goats were six months old. The vaccine was applied at this age because vaccination is indicated between three and six months of age, and this allowed us to follow the kids up to eleven months of age when they had already entered puberty. The first group was inoculated with the experimental strain: *B. melitensis* Rev 1 Δ eryCD. A fragment of 1158 bp was extracted from *Brucella abortus* S19, containing the Δ eryCD deletion, and was obtained by PCR using the oligonucleotides EryCD.R (5'-AGGGCCTTTGCTGTCGTTTC-3') and EryCD.F (5'-CAATCCGCTGGTCAACCGCT-3') with *B. abortus* S19 DNA. Then, the fragment was cloned into pGEM[®] T-Easy (Promega), completely sequenced to check for the absence of unwanted mutations introduced during the PCR step, and, subsequently, recloned as a *NotI* fragment into the mobilizable suicide plasmid pJQ200ucl1²², to produce pJQ- Δ eryCD. pJQ- Δ eryCD was introduced into the *Escherichia coli* S17-1 (λ -pir) mobilizing strain²⁷, and used to construct a deletion mutant in *B. melitensis* Rev 1, following the protocol previously described²⁶.

The second group was inoculated with *B. melitensis* Rev 1. The Rev 1 vaccine was produced by PRONABIVE Mexico laboratories. The third group (controls) received a vaccination with sterile physiological saline solution. In the three groups, the application was subcutaneous². The dosage for both strains was 1×10^9 colony forming units (CFU)/ml, the CFU count was performed following the Miles and Misra method².

Serological study

Serum was obtained from blood samples taken from the jugular vein on day zero, i.e. before the vaccination or euthanasia. Sera were kept in sterilized 1.5 ml tubes and frozen at -20°C before utilization. The serum was used for a 3% card test diagnosis of *Brucella* antibodies⁹.

Bacteriological study

Euthanasia was conducted under the guidelines of NOM-033-SAG/ZOO¹⁶. The animals were sacrificed using a concealed plunger gun and subsequent slitting, as approved by the Institutional Subcommittee for the Care and Use of Experimental Animals Number MC-2018/1-8.

Inguinal lymph nodes, mesenteric nodes, mediastinum, spleen, testicles, epididymis, seminal vesicles, bulbourethral glands, and an ampoule were collected for bacteriological, histology, immunohistochemistry, and PCR studies. Subsequently, three animals of each experimental group were euthanized monthly for the next five months.

The samples of nodes, mesenteric, mediastinum, spleen, testicles, epididymis, seminal vesicles, bulbourethral glands, and the ampoule, were independently macerated in a sterile 2 ml saline solution.

The primary isolation of *Brucella* spp. was performed by inoculating the samples onto Farrell agar plates and incubating them for 10 days at 37°C with 10% CO_2 . The bacterial

Table 1 Results of the 3% card test from vaccination to the time of euthanasia.

Groups	Post-vaccination days					
	0	35	63	91	126	183
Negative control	0/15	0/15	0/12	0/9	0/6	0/3
Rev 1	0/15	14/15	10/12	7/9	0/6	0/3
Δ eryCD	0/15	15/15	11/12	8/9	5/6	0/3

cultures were discarded after 10 days of incubation if no growth was visible². The typical colonies of *Brucella* spp. were stored at -80°C for further studies.

Eight 10-fold dilutions were made from this solution, each of which was inoculated onto Farrell plates to count CFU/ml. Next, they were incubated for 10 days at 37°C in an aerobic environment².

Histopathology, immunohistochemistry, and PCR

For the histopathological analysis, the sample tissues were stained with eosin/hematoxylin (EH). Immunohistochemistry was used to analyze the testicles and epididymis obtained during this study³.

DNA was extracted from testicle and epididymis samples using the commercial QIAamp DNA Mini Kit following the manufacturer's instructions. PCR conditions were as described by Sangari and Agüero²⁵.

Statistical analyses

Differences in seroconverted animals between groups in each evaluation were compared using the Chi-square test, SPSS version 25.

Results

All animals in this study were seronegative before vaccination (day 0). In the first month of analysis, 14/15 animals from the *B. melitensis* Rev 1 group and 15/15 from the *B. melitensis* Rev 1 Δ eryCD group exhibited vaccine antibodies. When tracking humoral immunity, it was found that the negative control group remained negative throughout the study. At each evaluation, the animals in the *B. melitensis* Rev 1 Δ eryCD group showed the highest seropositivity. Moreover, 2/3 animals in this group maintained antibody levels for the longest period, up to 4 months after vaccination. On the other hand, only 1/3 animal in the *B. melitensis* Rev 1 group showed a persistent serological response for up to 3 months after vaccination (Table 1). However, no evidence of difference between vaccinated groups was found ($p \geq 0.05$).

Throughout the study, *B. melitensis* was never isolated from the organs of the negative control group or the *B. melitensis* Rev 1 Δ eryCD group. The only exception occurred three months after vaccination, when *B. melitensis* was successfully isolated from left testicle samples of the *B. melitensis* Rev 1 group. PCR studies showed negative results for all tested organs. *B. melitensis* DNA was not detected even in the sample that proved positive in bacteriology.

Histopathological examination of testicles and accessory glands proved that the vaccination did not generate any changes in the reproductive system. The analyzed samples exhibited no pathological changes, not even in the animal from which *B. melitensis* was isolated. Immunohistochemistry results were compatible with the histopathological study, since no *B. melitensis* antigens were found in the analyzed testicles and epididymides, including in the sample testing positive in the bacteriological study.

Discussion

To this day, vaccines are still the most successful treatment to prevent infectious diseases in animals and humans¹⁹. *B. melitensis* Rev 1 vaccination has been used effectively around the world to control brucellosis in female goats and domesticated sheep⁴. The protection conferred by the Rev 1 *B. melitensis* vaccine at a reduced dose was evaluated in goats immunized five years earlier. Sixteen goats vaccinated five years earlier with Rev 1 with 1×10^5 CFU and 5 unvaccinated goats were challenged with *B. melitensis* 16M with 4×10^5 CFU by the conjunctival route. After calving or aborting, the goats were sacrificed, and tissue samples were taken for the bacteriological study. The challenge strain was recovered in 12% of the animals in the vaccinated group and 80% in the control group⁸.

Moreover, there is a conception that males are not relevant for the epidemiology of brucellosis. It has been suggested that vaccination could cause arthritis in up to 13% of vaccinated animals, which would affect their ability to reproduce³². Few studies have been conducted on this subject. Rev 1 vaccine has not been evaluated in young male goats and applied at a complete dose^{5,10,12,13}.

There are two previous cases of vaccination in males. Aldomy et al.¹ vaccinated adult males with a reduced dose of Rev 1, monitoring the post-vaccination serological response for up to 24 weeks, where the card test gave positive results throughout the study¹. Although they did not carry out a bacteriological study, their results showed longer persistence than those shown in our study, where the positive response to Rev 1 at a complete dose lasted for thirteen weeks.

In another study five adult domestic alpine goats and five mountain goats (*Capra ibex*) received complete dose conjunctival vaccination of Rev 1 vaccine, collecting sera and urethral swabs for serological and bacteriological monitoring. No clinical signs or specific Brucellosis lesions were observed, and, in the bacteriological study two male ibex presented urogenital excretion at 20 or 45 days post-vaccination. At sacrifice, isolates with a higher bacterial load were obtained at 45 days from ibex compared to domestic goats, while the levels remained between moderate and low when the animals were sacrificed 90 days post-vaccination²¹. This large number of isolates of the Rev 1 vaccine strain can be attributed to the use in this experiment of a complete Rev 1 dose, which is not recommended for adult animals. For this same reason a comparison with our study cannot be made, as we vaccinated kids with the complete dose of Rev 1 only recommended for young animals.

In the present study, male goats were vaccinated with a complete dose for the first time with the experimental

strain *B. melitensis* Rev 1 Δ eryCD and the strain *B. melitensis* Rev 1. The results in the serological follow-up using the card test were very similar in both vaccinated groups during the 3–4 months after vaccination. These data differ from other studies in which *B. melitensis* Rev 1 vaccination has been reported to generate a year-long humoral immunity in goats and sheep^{10,11}. This immune response is considered a problem because it interferes with the serological diagnosis after vaccination¹².

Nevertheless, protection against *Brucella* spp. requires a Th1 type immunological response¹⁸. Therefore, the cellular response should be studied, regardless of the short duration of the serological response. Moreover, the protection conferred by the studied strains should be further studied.

With regard to bacterial colonization, it is believed that *B. melitensis* Rev 1 shows tropism for the reproductive organs; however, the present study showed that none of the strains exhibited this characteristic. Only one of the 15 animals vaccinated with *B. melitensis* Rev 1 exhibited colonization of a testicle. In contrast, the strain could not be isolated from any of the animals vaccinated with *B. melitensis* Rev 1 Δ eryCD. Tolari and Salvi³⁰ isolated *Brucella* spp. from a goat that was ill with bilateral orchitis and had been immunized against *B. melitensis* Rev 1. These are the only studies in which positive isolation has been reported. Most studies have found that *Brucella* does not colonize the reproductive tract^{5,11–13}. In line with these studies, the present research has demonstrated relatively low tropism and colonization of the reproductive organs by both strains. In the case of *B. melitensis* Rev 1 Δ eryC, no tropism or colonization were identified. Therefore, this strain could represent a viable option to protect male goats against the illness and avoid the risk of infection.

In this study, the PCR was unable to detect *B. melitensis* DNA in a bacteriologically positive sample. The PCR only gave positive results when it was directly applied to the isolate. Nevertheless, this is not the first case in which bacteriology is more sensitive than PCR. A study by Buyukcangaz et al.⁶ on ruminant fetus organs, found that the sensitivity of PCR to detect *Brucella* spp. was 83% compared to bacteriology. Several reasons have been proposed to explain the low sensitivity of PCR in this context. These are, first, the presence of enzyme inhibitors in organ macerations⁶; second, the low number of bacteria in the samples⁶; and finally, the variations in specificity and sensitivity of the PCR method based on the laboratory characteristics²⁹, among other possible reasons. Although the present study expected higher sensitivity from the PCR, the results are similar to those in the previously mentioned studies. This could be related to some factors specified earlier, such as tissue contamination or the fact that it is a biological sample, without discarding the possibility of low quantities of bacteria in the sample.

Histopathological results showed that neither of the strains caused any detectable damage to the reproductive system. These results agree with the observations made by Blasco⁵, in which animals vaccinated with *B. melitensis* Rev 1 showed no lesions whatsoever. In those cases that did present some type of damage, it was not possible to isolate the strain; the challenging strain was isolated instead. In the present study, the *B. melitensis* Rev 1 strain was not isolated from most animals, which directly influenced the absence of injuries. In the case of the positive male goat, it is possible

that the infection was just starting to settle, therefore there were no evident alterations. The detection of *B. melitensis* in testicle and epididymis samples by immunohistochemical analysis was negative in all cases, even in the bacteriologically positive testicle sample. These negative results could be attributed to the fact that tissues colonized by bacteria were not included in the sample²⁸, or because the number of bacteria in the tissue was low¹⁴.

The present study shows that vaccination with *B. melitensis* strains Rev 1 Δ eryCD and Rev 1 causes no damage to the reproductive system of male goats. Strain *B. melitensis* Rev 1 Δ eryCD displayed reduced capacity to colonize the reproductive tract compared to strain Rev 1, due to its limited catabolic action toward erythritol. Results indicate that these strains do not represent a risk of infection to male goats when used for immunization against brucellosis, an important advantage when considering vaccination in controlled zones or with a high prevalence of the disease.

Further studies are needed regarding antibody permanence to prevent diagnostic misconceptions, and concerning long-term seminal quality in immunized animals to determine if fertility is affected.

Conflict of interest

The authors declare no conflicts of interest.

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References

- Aldomy F, Alkhalaf M, Younis IB. Immune responses of goats (Shami breed) to vaccination with a full, reduced and conjunctival dose of brucevac (*Brucella melitensis* Rev.1) vaccine. *Pak Vet J.* 2009;29:149–53.
- Alton G, Jones M, Pietz DE. Métodos bacteriológicos. Las técnicas de laboratorio en la brucelosis. World Health Organization, Food and Agriculture Organization of the United Nations; 1976.
- Beena V, Pawaiya RVS, Gururaj K, Singh DD, Mishra AK, Gangwar NK, Gupta VK, Singh R, Sharma AK, Karikalan M, Kumar A. Molecular etiopathology of naturally occurring reproductive diseases. *Vet World.* 2017;10:964–72.
- Blasco JM. A review of the use of *B. melitensis* Rev-1 vaccine in adult sheep and goats. *Prev Vet Med.* 1997;31:275–81.
- Blasco JM, Marín CM, Barberán M, Moriyón I, Díaz R. Immunization with *Brucella melitensis* Rev 1 against *Brucella ovis* infection of rams. *Vet Microbiol.* 1987;14:381–92.
- Buyukcangaz E, Sen A, Carli KT, Kahya S. Comparison of direct culture versus PCR for the detection of *Brucella* in aborted fetuses of cattle and sheep in Turkey. *Vet Rec.* 2011;168:430.
- Croch D, Elberg SS. Response of the vaccine strain of *Brucella melitensis* Rev 1 to erythritol. *J Bacteriol.* 1967;94:1793–4.
- Díaz-Aparicio E, Hernández L, Suárez-Güemes F. Protection against brucellosis in goats, five years after vaccination with reduced-dose *Brucella melitensis* Rev 1 vaccine. *Trop Anim Health Prod.* 2004;36:117–21.
- Díaz-Aparicio E, Marín C, Alonso-Urmeneta B, Aragón V, Pérez-Ortiz S, Pardo M, Blasco JM, Díaz R, Moriyón I. Evaluation of serological tests for diagnosis of *Brucella melitensis* infection of goats. *J Clin Microbiol.* 1994;32:1159–65.
- Fensterbank R, Pardon P, Marly J. Efficacy of *Brucella melitensis* Rev 1 vaccine against *Brucella ovis* infection in rams. *Ann Rech Vet.* 1982;13:185–90.
- Fensterbank R, Pardon P, Marly J. Comparison between the subcutaneous and conjunctival route of vaccination with Rev 1 strain against *Brucella melitensis* infection in ewes. *Ann Rech Vet.* 1982;13:295–301.
- García-Carrillo C. Protection of rams against *Brucella ovis* infection by *Brucella melitensis* Rev 1 vaccine. *Zentralbl Vet Med B.* 1981;28:425–31.
- Marín CM, Barberán M, Jiménez de Bagués MP, Blasco JM. Comparison of subcutaneous and conjunctival routes of Rev 1 vaccination for the prophylaxis of *Brucella ovis* infection in rams. *Res Vet Sci.* 1990;48:209–15.
- Meador VP, Tabatabai LB, Hagemoser WA, Deyoe BL. Identification of *Brucella abortus* in formalin-fixed, paraffin-embedded tissues of cows, goats, and mice with avidin-biotin-peroxidase complex immunoenzymatic staining technique. *Am J Vet Res.* 1986;47:2147–50.
- Muñoz PM, de Miguel MJ, Grilló MJ, Marín CM, Barberán M, Blasco JM. Immunopathological responses and kinetics of *Brucella melitensis* Rev 1 infection after subcutaneous or conjunctival vaccination in rams. *Vaccine.* 2008;19:2562–9.
- NOM-033-SAG/ZOO. Métodos para dar muerte a los animales domésticos y silvestres; 2014. https://www.gob.mx/cms/uploads/attachment/file/133499/4.NORMA_OFICIAL_MEXICANA_NOM-033-SAG-ZOO-2014.pdf
- NOM-041-ZOO. Campaña Nacional contra la Brucelosis en los animales; 1995. <https://www.gob.mx/cms/uploads/attachment/file/106184/NOM-041-ZOO-1995.pdf> [accessed 06.10.20].
- Oliveira SC, Splitter GA. CD8 type 1 CD4hi CD4RBlo T lymphocytes control intracellular *Brucella abortus* infection as demonstrated in major histocompatibility complex class I- and class II-deficient mice. *Eur J Immunol.* 1995;25:2551–7.
- Oñate AA, Céspedes S, Cabrera A, Rivers R, González A, Muñoz C, Folch H, Andrews E. A DNA vaccine encoding Cu, Zn superoxide dismutase of *Brucella*. *Infect Immun.* 2003;71:4857–61.
- Palomares REG, Aguilar RF, Flores PC, Gómez NL, Gutiérrez HJL, Herrera LE, Limón GM, Morales AJF, Pastor LF, Díaz Aparicio E. Enfermedades infecciosas de relevancia en la producción caprina, historia, retos y perspectivas. *Rev Mex Cienc Pecu.* 2021;12:205–23.
- Ponsart C, Riou M, Locatelli Y, Jacques I, Fadeau A, Jay M, Simon R, Perrot L, Freddi L, Breton S, Chaumeil T, Blanc B, Ortiz K, Vion C, Rioult D, Quéméré E, Sarradin P, Chollet JY, Garin-Bastuji B, Rossi S. *Brucella melitensis* Rev.1 vaccination generates a higher shedding risk of the vaccine strain in Alpine ibex (*Capra ibex*) compared to the domestic goat (*Capra hircus*). *Vet Res.* 2019;27:100.
- Quandt J, Hynes MF. Versatile suicide vectors which allow direct selection for gene replacement in gram-negative bacteria. *Gene.* 1993;15:15–21.
- Rossetti CA, Arenas-Gamboa AM, Maurizio E. Caprine brucellosis: a historically neglected disease with significant impact on public health. *PLoS Negl Trop Dis.* 2017;11:e0005692.
- Rossetti CA, Maurizio E, Rossi UA. Comparative review of *Brucellosis* in small domestic ruminants. *Front Vet Sci.* 2022;12:887671.
- Sangari FJ, Agüero J. Identification of *Brucella abortus* B19 vaccine strain by the detection of DNA polymorphism at the ery locus. *Vaccine.* 1994;12:435–8.
- Sangari FJ, Seoane A, Rodríguez MC, Agüero J, García Lobo JM. Characterization of the urease operon of *Brucella abortus*

- and assessment of its role in virulence of the bacterium. *Infect Immun.* 2007;75:774–80.
27. Simon R, Priefer U, Pühler A. A broad host range mobilization system for *in vivo* genetic engineering, transposon mutagenesis in Gram negative bacteria. *Nat Biotechnol.* 1983;1:784–91.
 28. Sözmen M, Erginsoy SD, Genç O, Beytut E, Özcan K. Immunohistochemical and microbiological detection of *Brucella abortus* antigens in aborted bovine fetuses. *Acta Vet Brno.* 2004;73:465–72.
 29. Tabibnejad M, Alikhani MY, Arjomandzadegan M, Hashemi SH, Naseri Z. The optimization of molecular detection of clinical isolates of *Brucella* in blood cultures by *eryD* transcriptase gene for confirmation of culture-negative samples. *Iran Red Crescent Med J.* 2016;18:e23879.
 30. Tolari F, Salvi G. Segnalazione di un caso di orchite bilaterale in un capretto in seguito a vaccinazione con Rev 1. Estrato dagli *Anm Faculta Med Vet.* 1980;33:87–91.
 31. Tosser-Klopp G, Bardou P, Bouchez O, Cabau C, Crooijmans R, Dong Y, Donnadieu-Tonon C, Eggen A, Heuven HC, Jamli S, Jiken AJ, Klop C, Lawley CT, McEwan J, Martin P, Moreno CR, Mulsant P, Nabihoudine I, Pailhoux E, Palhière I, Rupp R, Sarry J, Sayre BL, Tircazes A, Wan W, Wang W, Zhang W, International Goat Genome Consortium. Design and characterization of a 52K SNP chip for goats. *PLoS One.* 2014;22:e86227.
 32. West DM, Johnstone AC, Bruere AN, Chapman HM. Epiphysitis in rams following vaccination against *Brucella ovis* infection. *N Z Vet J.* 1978;26:133–4.