

Review

Silver jubilee: 25 years of the first demonstration of the direct effect of phosphate on the parathyroid cell

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ABSTRACT

Although phosphorus is an essential element for life, it is not found in nature in its native state but rather combined in the form of inorganic phosphates (PO_4^{3-}), with tightly regulated plasma levels that are associated with deleterious effects and mortality when these are out of bounds. The growing interest in the accumulation of PO_4^{3-} in human pathophysiology originated in its attributed role in the pathogenesis of secondary hyperparathyroidism (SHPT) in chronic kidney disease. In this article, we review the mechanisms by which this effect was justified and we commemorate the important contribution of a Spanish group led by Dr. M. Rodríguez, just 25 years ago, when they first demonstrated the direct effect of PO_4^{3-} on the regulation of the synthesis and secretion of parathyroid hormone by maintaining the structural integrity of the parathyroid glands in their original experimental model. In addition to demonstrating the importance of arachidonic acid (AA) and the phospholipase A2-AA pathway as a mediator of parathyroid gland response, these findings were predecessors of the recent description of the important role of PO_4^{3-} on the activity of the calcium sensor-receptor, and also fueled various lines of research on the importance of PO_4^{3-} overload not only for the pathophysiology of SHPT but also in its systemic pathogenic role.

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Bodas de plata: 25 años de la primera demostración del efecto directo del fósforo en la célula paratiroidea

RESUMEN

Aunque el fósforo es un elemento indispensable para la vida, en la naturaleza no se encuentra en estado nativo sino combinado en forma de fosfatos inorgánicos (PO_4^{3-}), con niveles plasmáticos estrechamente regulados que se asocian a efectos deletéreos y mortalidad cuando estos se encuentran fuera de la normalidad. El interés creciente sobre el acúmulo de PO_4^{3-} en la fisiopatología humana se originó en el papel que se le atribuyó en la patogenia del hiperparatiroidismo secundario a la enfermedad renal crónica. En este artículo revisamos los mecanismos por los cuales se justificaba dicho efecto y conmemoramos la importante contribución de un grupo español liderado por el Dr. M. Rodríguez, ahora hace justo 25 años, cuando demostraron por primera vez el efecto directo del PO_4^{3-} sobre la regulación de la síntesis y secreción de hormona paratiroidea, manteniendo la integridad estructural de las glándulas paratiroides en su nuevo modelo experimental. Además de demostrar la importancia del ácido araquidónico (AA) y la vía de fosfolipasa A2-AA como mediadora de respuestas en la glándula paratiroidea, estos hallazgos fueron predecesores de la reciente descripción del importante papel del PO_4^{3-} sobre la actividad del receptor-sensor de calcio y alimentaron asimismo diversas líneas de investigación sobre la importancia de la sobrecarga de PO_4^{3-} no sólo en la fisiopatología del HPTS sino también en su papel patogénico sistémico.

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Palabras clave:

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Fosfato

Hormona paratiroidea

PTH

Receptor de la PTH

CKD-MBD

Paratiroides

Introduction

Phosphorus (P), (in Greek *phos phorus* or carrier of light), is an essential element (atomic number 15, molecular weight 30.9 u) found in all living organisms. It has a dual function: (a) structural; in nucleic acids (DNA and RNA), cell membranes (phospholipids) and in the mineral phase of bone (together with calcium it forms hydroxyapatite crystals); (b) regulatory; constituting the main energy intermediate in cellular processes such as metabolism and activation of proteins (phosphorylation), involved in the oxygen dissociation curve of hemoglobin (2,3-diphosphoglycerate) and in cell signaling processes (second messengers such as cyclic AMP and GMP). In nature, including living organisms, phosphorus is not found in its native state, but mainly combined in the form of various non-flammable inorganic phosphates (PO_4^{3-}) (Fig. 1), this being the form in which phosphorus levels in the body are measured. Its biological importance determines that PO_4^{3-} levels must be strictly controlled within an optimal physiological range and that outside of these levels it is associated with deleterious effects. Thus, multiple epidemiological studies show not only that iPO_4^{3-} deficiency is a cause of pathology, but also elevated PO_4^{3-} levels (even in the normal ranges) have been independently associated with cardiovascular disease and mortality in the general population and, especially in patients with chronic kidney disease (CKD), whether on dialysis or not.^{1–7}

Considering that kidneys play a fundamental role in the regulation of PO_4^{3-} , alterations in its homeostasis in CKD are to be expected because the balance depends on its intake and excretion. The intra- and extracellular accumulation of PO_4^{3-} plays an essential role not only in the pathogenesis of secondary hyperparathyroidism (SHPT) and the chronic

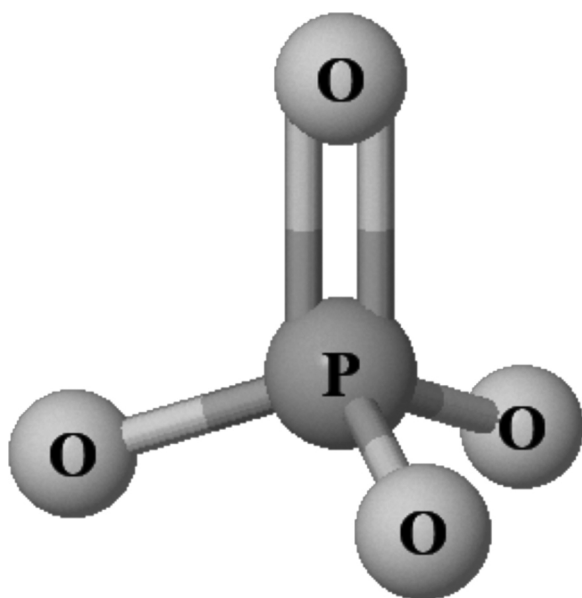


Fig. 1 – Molecular structure of the phosphate anion (esters of phosphoric acid): composed of a phosphorus atom and oxygen atoms in a tetrahedral shape. Made with the JSME Molecular Editor (<http://biomodel.uah.es/en/DIY/JSME/draw.en.htm>).

kidney disease-mineral and bone disorder (CKD-MBD) complex, but also, directly or indirectly, it can be considered as a (non-classical) risk factor for cardiovascular and even infectious morbidity and mortality, through various mechanisms including inflammation, oxidative stress, vascular and valvular calcifications or myocardial fibrosis. It also seems to contribute to cellular dysfunction of the immune system, arteriosclerosis-atheromatosis, and even accelerated aging in CKD patients.^{1,8–10} Recently, the role of PO_4^{3-} and calcium phosphate microcrystals in the renal tubular lumen have also become relevant, not only as a secondary effect but also as a causal factor in the progression of CKD itself.^{11,12}

However, the growing interest in PO_4^{3-} accumulation in human pathophysiology was originated by the role attributed to PO_4^{3-} in the aforementioned pathogenesis of SHPT. Therefore, the objective of this article is to review the mechanisms by which this effect was justified, and commemorate the important contribution of a Spanish group to the first demonstration in experimental studies of the *direct effect* of PO_4^{3-} on the regulation of synthesis and secretion of parathyroid hormone (PTH) in the parathyroid glands, just 25 years ago now.¹³

Phosphorus in the pathogenesis of secondary hyperparathyroidism: indirect mechanisms

For many years there has been a profound, almost bipolar, debate about whether the retention of PO_4^{3-} or the decrease in the synthesis of calcitriol (1,25- $[\text{OH}]_2$ -vitamin D), both present in patients with CKD, were the initiating or most important factors in the pathogenesis of SHPT.^{14–16} Exactly five decades ago now (golden anniversary), Slatopolsky et al.¹⁴ observed a significant increase in PTH in an experimental model in dogs with CKD fed a high PO_4^{3-} diet. The interpretation of these findings was that a transient increase in postprandial PO_4^{3-} would decrease ionized calcium in the blood, and this decrease in calcium would be what would stimulate PTH secretion. The increase in PTH would not only normalize calcium levels, but would also decrease tubular phosphorus reabsorption, with the consequent increase in phosphaturia, thus normalizing the serum levels of both ions at the expense of elevated serum PTH. Subsequent decreases in glomerular filtration (GFR) would lead to a progressive increase in the PTH levels necessary to normalize calcium and PO_4^{3-} . These observations constituted the “*trade-off hypothesis*” (or PTH elevation –and its consequent deleterious effects– at the expense of trying to maintaining calcium and PO_4^{3-} homeostasis)^{17,18} (Fig. 2). These same authors demonstrated in another experimental study that the proportional reduction in PO_4^{3-} intake adjusted to the gradual decrease in GFR was able to prevent SHPT, as evidenced by the presence of normal plasmatic levels of calcium, PO_4^{3-} , PTH and tubular phosphorus reabsorption as compared to controls.¹⁹ In this experimental model, the explanation for the observations made was based on the fact that the adaptation of the remaining nephrons to the CKD stage was necessary, in which these nephrons must respond to a greater fraction of renal excretion of PO_4^{3-} with a decrease of its tubular reabsorption (as usually happens in the unadjusted intake of PO_4^{3-} or other elements).²⁰ This hypothesis

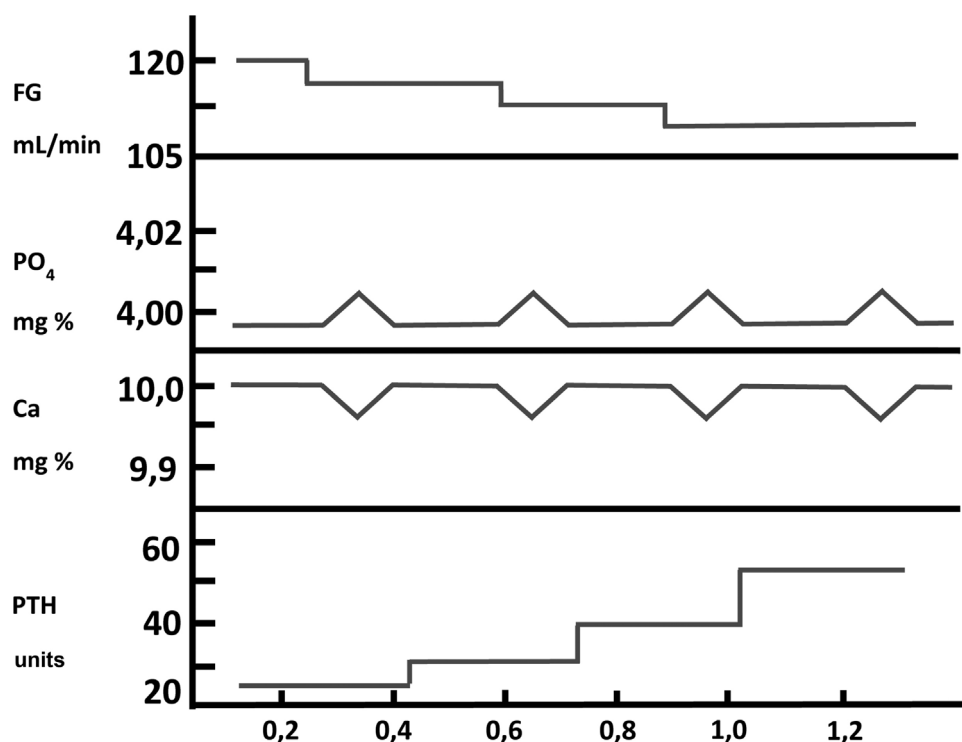


Fig. 2 – Pathogenesis of secondary hyperparathyroidism in chronic kidney disease. Representation of the « trade-off hypothesis ». (Adapted from Bricker et al.¹⁸).

was postulated by Bricker under the so-called «intact nephron hypothesis»²⁰ and the findings then suggested that a constant intake of PO_4^{3-} in the face of a diminished nephron population (CKD) would play an important role in the pathogenesis of SHPT.^{20,21}

Although in the aforementioned study¹⁹ the reduction in PO_4^{3-} prevented SHPT, the possibility of the contribution of PTH inhibition by an increased calcitriol synthesis induced by the low PO_4^{3-} diet could not be ruled out. This fact would be particularly relevant, especially in the early stages of kidney damage, as demonstrated by Portale et al.¹⁵ years later in children with moderate CKD. PO_4^{3-} is a known inhibitor of 1 α -hydroxylase (CYP27B1) and in these children it was shown that a diet high in PO_4^{3-} stimulated PTH and decreased calcitriol levels, whereas a diet low in PO_4^{3-} stimulated calcitriol with a secondary decrease in PTH. In addition, it is common for a diet low in PO_4^{3-} to be associated with an increase in serum calcium due to the greater intestinal absorption of calcium secondary to the increase in calcitriol, so that this increase in serum calcium could explain, at least in part, the decrease in PTH. Thus, Llach and Massry¹⁶ demonstrated in four patients with moderate CKD that PO_4^{3-} restriction favored the action and production of calcitriol, modifying not only intestinal calcium absorption but also the calcemic response to PTH, thus suggesting an important role of altered vitamin D metabolism in the pathogenesis of SHPT in CKD.

In those days, the «so called «decreased calcemic response or skeletal resistance to the action of PTH (endogenous and exogenous) was another *indirect mechanism* described by which PO_4^{3-} retention and/or hyperphosphatemia could contribute to the development and progression of SHPT in CKD,^{22,23} and

this effect has been widely studied by Rodriguez et al.²⁴⁻²⁷ Currently called « hyporesponsiveness» to PTH,²⁸ various studies have shown the relative importance of PO_4^{3-} retention on the resistance to the action of PTH, among other factors.²⁴⁻³⁰ As reviewed in a recent article in this journal,³⁰ the existence of a hyporesponse to the action of PTH, through various mechanisms, among which it stands out the retention of PO_4^{3-} , demands a greater synthesis and secretion of PTH to maintain mineral homeostatic balance.

Phosphorus in the pathogenesis of secondary hyperparathyroidism: direct mechanism

Until now, the mechanisms described in this review on the role of PO_4^{3-} in the pathogenesis of SHPT have been *indirect* (through a decrease in ionic calcium, a decrease in calcitriol synthesis or multifactorial hyporesponsiveness to the actions of PTH in CKD). It is well known that both the decrease in extracellular calcium concentration and the decrease in calcitriol levels have a direct stimulatory effect on PTH synthesis and parathyroid cell proliferation, mediated by their respective receptors (the calcium sensing receptor [CaSR] and vitamin D receptor [VDR]).³¹⁻³⁴ In fact, Sherwood et al.³⁵ failed to find evidence of a *direct effect* of PO_4^{3-} on the regulation of PTH *in vivo*.

However, López-Hilker et al.,³⁶ in another experimental study in dogs with CKD on PO_4^{3-} and also calcium restriction in order to prevent hypercalcemia, observed that the serum PTH level decreased even without changes in calcium and calcitriol. These findings suggested that the control

of SHPT was independent of plasma calcium and calcitriol levels. Years later, in an experimental model in rats, Yi et al.³⁷ also found evidence of the existence of calcium and calcitriol-independent mechanisms to control PTH and, therefore, a possible *direct effect* of PO_4^{3-} on parathyroid function.

On the other hand, since it is difficult to ignore the direct effect of calcitriol on the decrease in PTH synthesis and secretion when a low PO_4^{3-} diet is prescribed, Kilav et al.³⁸ also demonstrated that second-generation vitamin D-deficient rats fed a diet low in PO_4^{3-} and calcium had hypophosphatemia associated with low PTH mRNA levels in the absence of hypercalcemia or increased calcitriol levels, suggesting a non-transcriptional effect of PO_4^{3-} , in contrast to the direct effects of calcitriol by decreasing the transcription of the pre-pro-PTH gene. It has subsequently been shown that this non-transcriptional effect of PO_4^{3-} is actually *post-transcriptional*, involving the binding of transactivating proteins (proteins that act on trans or adenosine-uridine-rich binding factor [AUF1]) to cis domains (cis element) located at the 3' untranslated region of PTH mRNA, orchestrated by Pin1 isomerase, ultimately leading to increased PTH mRNA stability.³⁹⁻⁴²

Finally, in a preliminary work, Hernández et al.⁴³ had also described in a preliminary work the effects of a diet high in PO_4^{3-} in an *in vivo* rat model and found, compared to rats that ingested a standard PO_4^{3-} diet, a significant increase in the levels of PTH associated with a rapid increase in PTH mRNA expression without affecting calcium or calcitriol levels. As they subsequently demonstrated, this observation was not accompanied by changes in the expression of VDR or RSCa,⁴³⁻⁴⁵ suggesting that an oral PO_4^{3-} load could *directly* stimulate PTH synthesis and that it was not mediated by decreased expression of the calcium or calcitriol receptors.

Then, using a completely original experimental model, based on the maintenance of the structural integrity of the parathyroid gland, a Spanish group led by Dr. M. Rodríguez described for the first time *in vitro* and in an unquestionable manner the *direct effect* of PO_4^{3-} on PTH secretion (IX Latin American Congress of Nephrology in San Juan, Puerto Rico, October 20-23, [1994] and the XIIIth International Congress of Nephrology in Madrid, July 2-6, [1995]). Thus, Almadén et al.¹³ published in 1996 their work in which they used whole fresh rat parathyroid glands that were incubated with different concentrations of PO_4^{3-} (1, 2, 3, and 4 mM) and were subsequently exposed to different calcium concentrations in a range from 0.4 to 1.35 mM. The authors found that at a calcium concentration of 1.25 mM, PTH secretion was similar with PO_4^{3-} concentrations of 1 and 2 mM; however, PO_4^{3-} concentrations of 3 and 4 mM produced an increase in PTH secretion three and four times greater, respectively, as compared to PO_4^{3-} of 1 mM (Fig. 3). Likewise, in concentrations of 1 or 2 mM of PO_4^{3-} an increase in calcium concentration from 0.6 to 1.35 mM reduced PTH secretion to 37%; whereas in PO_4^{3-} of 4 mM the same increase in calcium concentration inhibited PTH secretion only to 75% (Fig. 3). This pioneering demonstration of the essential role of the structural integrity of the parathyroid gland and the maintenance of cell-cell contacts was a fundamental starting point to accelerate the progress of knowledge on the impact of PO_4^{3-} in the pathogenesis of

SHPT in different laboratories around the world. In fact, until then, with the same objective of evaluating the possible direct effect of PO_4^{3-} on parathyroid function, dispersed parathyroid cells were used in *in vitro* studies showing a lower response to changes in extracellular calcium than in the intact parathyroid gland model used by Almadén et al.¹³ The fact is that in isolated (dispersed) parathyroid cells the response to changes in extracellular calcium is reduced due to a decreased expression of the RSCa.⁴⁶

In this seminal study by Almadén et al.,¹³ the authors also studied the intracellular signaling mechanisms that mediate the direct effect of PO_4^{3-} on PTH secretion. Thus, the addition of arachidonic acid (AA, a substrate that inhibits the intracellular signaling pathway), to the incubation medium of PO_4^{3-} 4 mM and calcium 1.35 mM reduced PTH secretion to 34.5%. The conclusion of the study was therefore that in this model using fresh whole rat parathyroid glands, the elevation of PO_4^{3-} *in vitro* directly increased PTH secretion by acting through the phospholipase A2 (PLA2)-AA pathway.

Subsequently, they also demonstrated *in vitro* that, in glands from patients with severe SHPT, the PTH secretion increased in response to PO_4^{3-} 3 and 4 mM as compared to 2 mM despite the presence of a high concentration of calcium in the medium; this effect was accompanied by an increase in pre-pro-PTH.⁴⁷ Likewise, this same group demonstrated that an *acute elevation* of serum PO_4^{3-} , without changes in the ionic calcium concentration, also stimulated PTH secretion *in vivo*.⁴⁸ They also showed the effect of high PO_4^{3-} concentrations on AA production in parathyroid tissue *in vitro*,⁴⁹ and the regulation of AA production by intracellular calcium,⁵⁰ showing that the stimulation of PTH secretion by high PO_4^{3-} levels could be prevented by increasing intracellular calcium levels. They also reviewed the importance of AA and the phospholipase A (2)-AA pathway as a mediator of responses in the parathyroid gland.⁵¹ In addition, it was shown that maintaining elevated serum PO_4^{3-} levels during the construction of calcium-PTH curves in hemodialysis (using high or free PO_4^{3-} dialysis fluid) partially prevented the inhibition of PTH secretion by calcium (normal or elevated).⁵²

On the other hand, from the laboratory of Slatopolsky et al. (J of Investigative Medicine, April [1995]), using the same experimental model of intact parathyroid glands as used by Almadén et al.,¹³ it was published in the same year, 1996, the effect of dietary PO_4^{3-} on PTH levels, PTH mRNA, and parathyroid hyperplasia in uremic and normal rats.⁵³ The authors observed that gland weight and serum PTH were similar in both groups exposed to a low PO_4^{3-} diet (0.2%), but there was a significant increase in serum PTH, gland weight, and DNA in the parathyroid glands from uremic rats fed a high PO_4^{3-} diet (0.8%) compared with uremic rats fed a low PO_4^{3-} diet. Additionally, they observed that the stimulatory effect of extracellular PO_4^{3-} on PTH production did not occur when protein synthesis was inhibited with cycloheximide, suggesting that the action of PO_4^{3-} on parathyroid cells required protein synthesis. Likewise, the growth rate of parathyroid cells was independent of calcium and calcitriol levels. Previously, Denda et al.⁵⁴ had already shown that the effect of PO_4^{3-} on the growth of parathyroid glands was very rapid in uremic rats, two months after the induction of renal failure, and that 90% of the growth occurred during the first three days, poten-

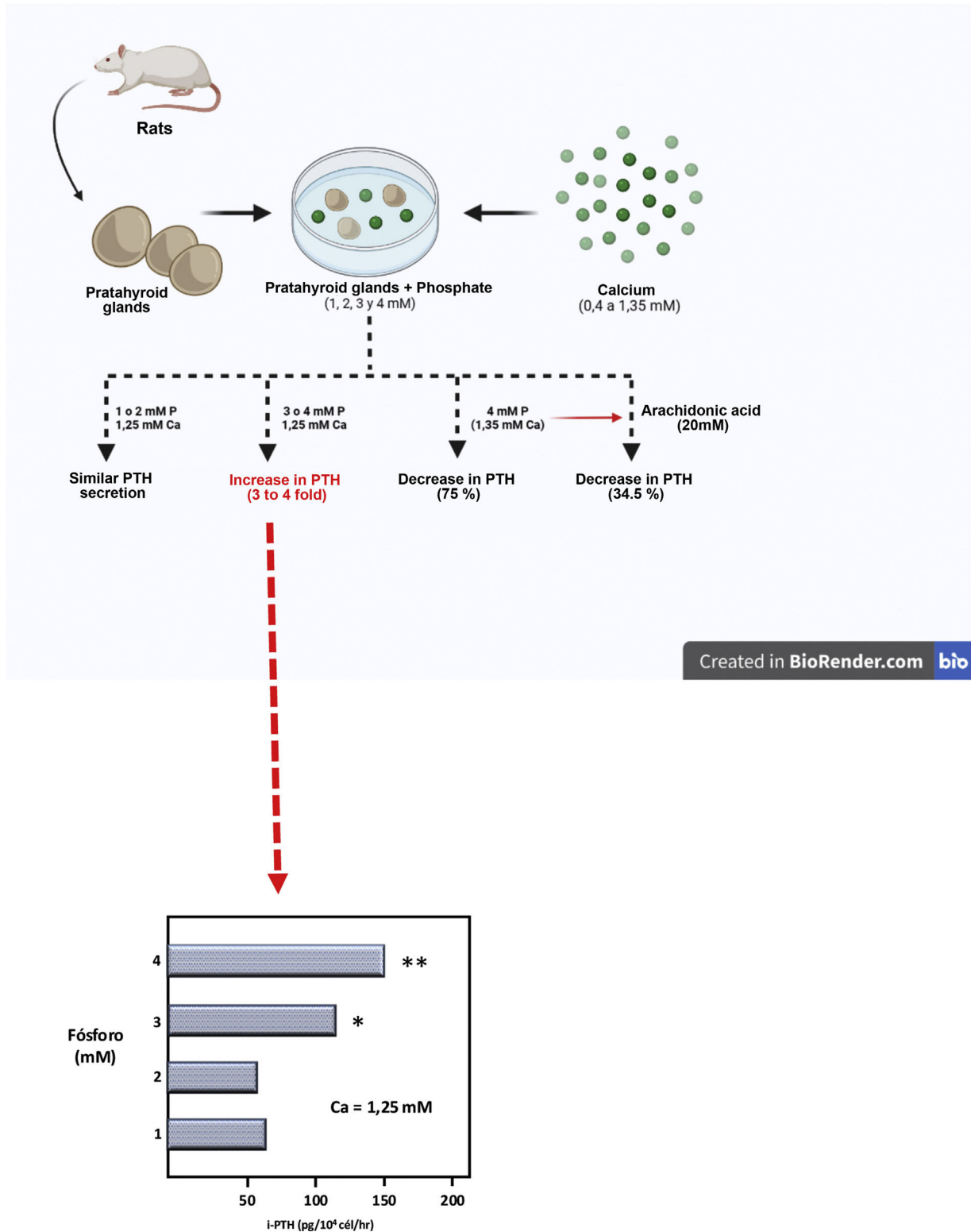


Fig. 3 – Theoretical representation of the model used by Almadén et al.¹³ for the demonstration of the direct effect of phosphorus (P), independent of calcium (Ca), on the stimulation of parathyroid hormone (PTH) secretion in the parathyroid gland. The glands were incubated for 1 h in a calcium concentration of 1.25 mM and a variable phosphorus concentration of 1, 2, 3 and 4 mM.

tially attributed to the participation of proto-oncogenes such as *c-fos*, *c-jun* and *PRAD-1*.

Interestingly, in the same year, acknowledging the work of the two groups led by doctors Rodríguez and Slatopolsky, and with the same objective of investigating the direct effect of PO_4^{3-} on parathyroid cells *in vitro*, Kjaerulff et al. from Dr Olgaard's Danish group,⁵⁵ used two types of bovine parathyroid tissue preparations: dispersed parathyroid cells and parathyroid tissue sections, both incubated for four hours in normal (1.0 mM) or high PO_4^{3-} (3.5 mM) and observed a significant increase in the release of PTH in the parathyroid tissue sections incubated in medium high in PO_4^{3-} but not in the preparation with dispersed cells, without observing changes in the "set-point" of calcium. The degree of stimulation of PTH release with high PO_4^{3-} in the medium was significantly higher compared with low calcium medium (0.8 mM), 172% above baseline (1.0 mM PO_4^{3-}) and 139% higher in a high calcium medium (1.8%). Their results also demonstrated that PO_4^{3-} directly stimulates PTH release in sections of bovine parathyroid glands and not in dispersed cell preparations, confirming that maintenance of normal parathyroid architecture is essential to reproduce the stimulatory effect of increasing PO_4^{3-} in PTH secretion.

Effect of phosphorus on the parathyroid glands

Under normal conditions, parathyroid cells are in a quiescent state and rarely undergo mitosis; however, it is well known that in CKD several factors such as hypocalcemia and calcitriol deficiency induce growth of the parathyroid glands, stimulating parathyroid cell proliferation of parathyroid cells, initially at the expense of cell hyperplasia and, consequently, the synthesis and secretion of PTH.^{56–59} We have already mentioned that a diet high in PO_4^{3-} , either by direct or indirect mechanisms, promotes parathyroid growth from an early stage, as demonstrated by Denda et al. in experimental studies in rats.⁵⁴ Conversely, a low PO_4^{3-} diet and calcitriol administration were also shown to prevent uremia-induced hyperplasia and PTH secretion. Thus, Dusso et al.⁶⁰ demonstrated in uremic rats that a diet low in PO_4^{3-} was able to prevent parathyroid gland hyperplasia by increasing p21 protein and mRNA in parathyroid tissue. The protein encoded by the p21 gene is an inhibitor of cyclin-dependent kinases (Cdk complexes) and, therefore, a regulator of the cell cycle. Likewise, this study also demonstrated that a diet high in PO_4^{3-} induced the expression of transforming growth factor- α (TGF- α) as an autocrine signal that stimulated parathyroid growth. Such elevations of TGF- α in the parathyroid gland induced by hyperphosphatemic diets and the consequent activation of the epidermal growth factor receptor (EGFR) were identified as the determinants of the decrease in vitamin D receptor (VDR) and the origin of resistance to control of HPTS with calcitriol or its analogues with the progression of CKD.⁶¹ It is also important to highlight that, as CKD progresses, not only VDR expression decreases, but also the expression of the CaSR and fibroblast growth factor receptor 23 (FGFR) during the evolution from *diffuse* hyperplasia to *nodular hyperplasia*.⁵⁹ More recently, it was also shown that a high demand for PTH secretion, promoted either by a diet very high in PO_4^{3-} or low in calcium, induced different

patterns of parathyroid hyperplasia in the absence of uremia, a situation that could be important in early stages of CKD.⁶²

It is now interesting to note that the regulation of PTH by PO_4^{3-} also involves certain *micro-RNA* (miRNA).^{63,64} The miRNAs are small non-coding RNAs with vital functions in homeostasis and development of the organism, and we know that the *Dicer enzyme* is involved in the final stage of miRNA processing. In this regard, for example, we have recently learned that parathyroid cell *Dicer*-specific *knockout mice* (*PT-Dicer* *-/-*) have normal serum PTH levels, but cannot increase PTH in hypocalcemia or renal failure, unlike controls.⁶⁴ Also, in addition to modulating PTH secretion, miRNAs are essential for keeping intact parathyroid glands. *Dicer knockout mice* do not express miRNA in parathyroid cells and lose their parathyroid glands after birth. This indicates that miRNAs are not essential for embryonic development of the parathyroid glands, but rather for their integrity during the postnatal period. In the absence of parathyroid glands, in adult *PT-Dicer* *-/-* mice, an additional source of PTH, other than thyroid cells or the thymus, contributes to the maintenance of normal serum PTH concentrations, has been demonstrated but cannot be stimulated by hypocalcemia or an uremic state.⁶⁴

Other aspects related to phosphorus overload in chronic kidney disease

It is not the purpose of this article to review the entire complex pathophysiology of SHPT but, as mentioned, to commemorate the 25th anniversary of the important discovery of the *direct effect* of PO_4^{3-} on the parathyroid gland, in which Spanish researchers played such an important role.^{13,53} However, it is necessary to mention the importance of the discovery of a *phostatonin*, fibroblast growth factor 23 (FGF23),^{65,66} a hormone produced by osteocytes and whose production is mainly stimulated by PO_4^{3-} overload, among other factors.^{67,68} Today we know that its elevation occurs from early stages of CKD, even before the elevation of serum PTH, which in turn also stimulates the production of FGF23.^{69–71} FGF23 not only has a phosphaturic action by inhibiting the expression of Na-Pi 2a and 2c channels at the renal tubular level (decreasing tubular reabsorption of PO_4^{3-} and thus increasing its urinary excretion) but also inhibits renal 1 α -hydroxylase (CYP27B1) (responsible for the synthesis of calcitriol) and stimulates 24-hydroxylase (CYP24A1) (increasing its catabolism).^{69,70} This new counterregulatory mechanism of PO_4^{3-} overload allows us to revisit the old, but still current, "trade-off" hypothesis^{17,21,70,72} and the importance of PO_4^{3-} in the pathogenesis of SHPT in CKD, by providing a new previously unknown mediator, the FGF23. This increase in FGF23 is explained at least in part by the reduced expression of its co-receptor *Klotho*, which is necessary to exert its action in target tissues (parathyroid, vascular, brain, renal tubules). This has been demonstrated in several studies and in the case of kidney tissue, it will lead to resistance to the action of FGF23.⁷³ It is known that the action of FGF23 is mediated by a canonical affinity between its receptor FGFR and its *Klotho* co-receptor,^{10,74} so that by decreasing *Klotho* production and/or FGFR expression as a consequence of CKD,^{73–76} it will cause another hormonal hyporesponse

(resistance), in addition to that of PTH or other hormones such as insulin or growth hormone.³⁰ This hyporesponse to FGF23 will lead to an additional increase in the levels of FGF23 required to exert its phosphaturic action in the presence of CKD and/or phosphorus overload, but the excess of FGF23 will produce deleterious secondary effects (such as another "trade-off", in this case at the expense of high FGF23).^{68,73,77–79} In fact, both the decrease in Klotho and the increase in FGF23 typical of CKD have been clearly associated with accelerated aging and disproportionately high mortality in CKD patients, especially in its advanced stages or on dialysis,^{2,10,80} turning CKD into an unfortunate human experimental model of senescence.^{8,9} Therefore, the intra- and extracellular retention of PO_4^{3-} constitutes one of the main stimuli for the synthesis and secretion of both hormones (PTH and FGF23), either directly or indirectly, with the purpose of increasing phosphaturia, among other effects. In addition, PTH and FGF23 act competitively in the enzymatic regulation of vitamin D production and catabolism. This complex hormonal interrelationship is exacerbated by the progressive loss of renal parenchyma as CKD progresses. Furthermore, the secondary reduction of klotho, the increase in PO_4^{3-} and the increase in FGF23 act as proinflammatory stimuli, being key elements in the inflammatory state consubstantial to CKD and involved in multiple deleterious effects associated with the loss of renal function. (development of anemia and resistance to erythropoiesis-stimulating agents, protein-energy malnutrition syndrome, endothelial dysfunction), as well as early and accelerated calcifications and atherosclerosis.^{81–83}

Corollary

Under normal conditions, homeostasis of extracellular PO_4^{3-} is coordinated between intestinal absorption, renal excretion, as well as its entrance and exit from bone.⁸⁴ Both parathyroid glands and bone detect increase in extracellular PO_4^{3-} and they react by increasing the levels of PTH and FGF23, respectively, with the purpose of increasing phosphaturia; however, the intrinsic molecular mechanism sensing the extracellular PO_4^{3-} is unknown. This is in contrast with what happens with the interactions of calcium or calcimimetics with the RSCa, vitamin D with the VDR and even with FGF23 and its own receptor.^{33,46,59,75} For many years, attempts have been made to find the PO_4^{3-} receptor in the parathyroid gland, trying to explain its direct effect, and even postulating the possibility of the existence of a transporter channel.

It is only recently that the importance of the CaSR in sensing the extracellular level of PO_4^{3-} has been demonstrated. The CaSR is found in the membranes of a variety of cells and belongs to the family of G protein-coupled receptors. It was already known that calcimimetics could overcome the stimulatory effect of increasing PO_4^{3-} levels on the secretion of PTH *in vitro* and *in vivo*.⁸⁵ Recently, Geng et al.⁸⁶ used an X-ray crystallography technique to study the three-dimensional structure of the outer domain of the RSCa in active and resting states. It was observed that calcium ions are the main activators of the RSCa but it requires the binding of amino acids in its active form. Likewise, the extracellular domain of the RSCa was also found to have four multivalent anion

binding sites occupied by PO_4^{3-} and SO_4^{2-} and that PO_4^{3-} ions kept stable the inactive form of the CaSR, thus promoting PTH secretion. Studies by Centeno et al.⁸⁷ recently showed in experimental murine models that the increase in PO_4^{3-} in pathophysiological concentrations of CKD inhibits the activity of the RSCa in an antagonistic, non-competitive way. Thus, these findings finally show us that the RSCa is also a sensor of PO_4^{3-} , explaining the intrinsic mechanism by which PO_4^{3-} directly stimulates PTH secretion and providing a mechanism by which elevated concentrations of PO_4^{3-} can exert direct effects on tissues expressing the RSCa.

In summary, PO_4^{3-} overload leads to the activation of various direct and indirect mechanisms aiming to maintain its homeostasis.⁸⁸ The presence of CKD creates at least two vicious cycles in CKD-MBD, the increase in PTH and in FGF23, with the consequent deleterious effects (double "trade-off" that not only affects bone but also the cardiovascular system),^{1,8,68} and these undesirable effects are the consequence of a failed attempt to normalize mineral metabolism. With the progression of CKD, there is an accumulation of PO_4^{3-} , and therefore it is important to especially restrict the sources of inorganic PO_4^{3-} to prevent the development of SHPT; the stimulation of FGF23, the inhibition of calcitriol synthesis and the resulting hypocalcemia, its negative effect on the calcemic response to the action of PTH or the stabilization of the inactive form of the RSCa. PO_4^{3-} clearly blocks all known counterregulatory mechanisms designed to maintain mineral metabolism homeostasis in healthy situations. In addition, this accumulation of PO_4^{3-} directly contributes to clear harmful effects mediated by multiple mechanisms that affect the cardiovascular system or the kidney itself^{1,8,12,89–92} and which seem to explain the very important association of PO_4^{3-} overload on morbidity and mortality in patients with CKD.

Conflict of interests

JB has received honoraria as a consultant, speaker, or travel support from Amgen, Abbvie, Sanofi, and Vifor-Fresenius-Renal Pharma. PU has received fees as a consultant or speaker from Amgen, Astellas, GSK, Hemotech, Leo-Pharma, Sanofi, and Vifor-Fresenius-Renal Pharma. AT has received consulting fees from Alnylan Pharm. JFNG has received honoraria as a consultant, speaker, or travel support from Abbvie, Amgen, Sanofi-Genzyme, Shire, and Vifor-Pharma. JF has received fees as a consultant from Vifor-Pharma.

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REFERENCES

1. Vervloet MG, Sezer S, Massy ZA, Johansson L, Cozzolino M, Fouque D, et al. The role of phosphate in kidney disease. *Nat Rev Nephrol.* 2017;13:27–38.

2. Gutiérrez OM, Mannstadt M, Isakova T, Rauh-Hain JA, Tamez H, Shah A, et al. Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. *N Engl J Med*. 2008;359:584-92.
3. Block GA, Klassen PS, Lazarus JM, Ofsthun N, Lowrie EG, Chertow GM. Mineral metabolism, mortality, and morbidity in maintenance hemodialysis. *J Am Soc Nephrol*. 2004;15:2208-18.
4. Palmer SC, Hayen A, Macaskill P, Pellegrini F, Craig JC, Elder GJ, et al. Serum levels of phosphorus, parathyroid hormone, and calcium and risks of death and cardiovascular disease in individuals with chronic kidney disease: a systematic review and meta-analysis. *JAMA*. 2011;305:1119-27.
5. Tonelli M, Sacks F, Pfeffer M, Gao Z, Curhan G, Cholesterol And Recurrent Events Trial Investigators. Relation between serum phosphate level and cardiovascular event rate in people with coronary disease. *Circulation*. 2005;112:2627-33.
6. Lloret MJ, Bover J, DaSilva I, Furlano M, Ruiz-García C, Ayasreh N. Papel del fósforo en la enfermedad renal crónica. *Nefrología Sup Ext*. 2013;4(2):2-10.
7. Danese MD, Belozeroff V, Smirnakis K, Rothman KJ. Consistent control of mineral and bone disorder in incident hemodialysis patients. *Clin J Am Soc Nephrol*. 2008;3:1423-9.
8. Covic A, Vervloet M, Massy ZA, Torres PU, Goldsmith D, Brandenburg V, et al. Bone and mineral disorders in chronic kidney disease: implications for cardiovascular health and ageing in the general population. *Lancet Diabetes Endocrinol*. 2018;6:319-31.
9. Kuro OM. Phosphate as a pathogen of arteriosclerosis and aging. *J Atheroscler Thromb*. 2021;28:203-13.
10. Kuro OM. Klotho and calciprotein particles as therapeutic targets against accelerated ageing. *Clin Sci*. 2021;135:1915-27.
11. Shiizaki K, Tsubouchi A, Miura Y, Seo K, Kuchimaru T, Hayashi H, et al. Calcium phosphate microcrystals in the renal tubular fluid accelerate chronic kidney disease progression. *J Clin Invest*. 2021:131.
12. Letavernier E, Drüeke TB. Kidney toxicity of phosphate: is that crystal clear yet? *Kidney Int*. 2021;100:1155-7.
13. Almaden Y, Canalejo A, Hernandez A, Ballesteros E, Garcia-Navarro S, Torres A, et al. Direct effect of phosphorus on PTH secretion from whole rat parathyroid glands in vitro. *J Bone Miner Res*. 1996;11:970-6.
14. Slatopolsky E, Caglar S, Pennell JP, Taggart DD, Canterbury JM, Reiss E, et al. On the pathogenesis of hyperparathyroidism in chronic experimental renal insufficiency in the dog. *J Clin Invest*. 1971;50:492-9.
15. Portale AA, Booth BE, Halloran BP, Morris RC Jr. Effect of dietary phosphorus on circulating concentrations of 1,25-dihydroxyvitamin D and immunoreactive parathyroid hormone in children with moderate renal insufficiency. *J Clin Invest*. 1984;73:1580-9.
16. Llach F, Massry SG. On the mechanism of secondary hyperparathyroidism in moderate renal insufficiency. *J Clin Endocrinol Metab*. 1985;61:601-6.
17. Bricker NS. On the pathogenesis of the uremic state. An exposition of the "trade-off hypothesis." *N Engl J Med*. 1972;286:1093-9.
18. Bricker NS, Slatopolsky E, Reiss E, Avioli LV. Calcium, phosphorus and bone in renal disease and transplantation. *Arch Intern Med*. 1969;123:543-53.
19. Slatopolsky E, Caglar S, Gradowska L, Canterbury J, Reiss E, Bricker NS. On the prevention of secondary hyperparathyroidism in experimental chronic renal disease using "proportional reduction" of dietary phosphorus intake. *Kidney Int*. 1972;2:147-51.
20. Bricker NS, Morrin PA, Kime SW Jr. The pathologic physiology of chronic Bright's disease. An exposition of the "intact nephron hypothesis." *Am J Med*. 1960;28:77-98.
21. Slatopolsky E. The intact nephron hypothesis: the concept and its implications for phosphate management in CKD-related mineral and bone disorder. *Kidney Int Suppl*. 2011:S3-8.
22. Evanson JM. The response to the infusion of parathyroid extract in hypocalcaemic states. *Clin Sci*. 1966;31:63-75.
23. Llach F, Massry SG, Singer FR, Kurokawa K, Kaye JH, Coburn JW. Skeletal resistance to endogenous parathyroid hormone in patients with early renal failure. A possible cause for secondary hyperparathyroidism. *J Clin Endocrinol Metab*. 1975;41:339-45.
24. Rodriguez M, Felsenfeld AJ, Llach F. Calcemic response to parathyroid hormone in renal failure: role of calcitriol and the effect of parathyroidectomy. *Kidney Int*. 1991;40:1063-8.
25. Rodriguez M, Martin-Malo A, Martinez ME, Torres A, Felsenfeld AJ, Llach F. Calcemic response to parathyroid hormone in renal failure: role of phosphorus and its effect on calcitriol. *Kidney Int*. 1991;40:1055-62.
26. Bover J, Jara A, Trinidad P, Rodriguez M, Martin-Malo A, Felsenfeld AJ. The calcemic response to PTH in the rat: effect of elevated PTH levels and uremia. *Kidney Int*. 1994;46:310-7.
27. Bover J, Rodriguez M, Trinidad P, Jara A, Martinez ME, Machado L, et al. Factors in the development of secondary hyperparathyroidism during graded renal failure in the rat. *Kidney Int*. 1994;45:953-61.
28. Evenepoel P, Bover J, Ureña Torres P. Parathyroid hormone metabolism and signaling in health and chronic kidney disease. *Kidney Int*. 2016;90:1184-90.
29. Ureña P, Kubrusly M, Mannstadt M, Hruby M, Trinh MM, Silve C, et al. The renal PTH/PTHrP receptor is down-regulated in rats with chronic renal failure. *Kidney Int*. 1994;45(2):605-11.
30. Hiporrespuesta o resistencia a la acción de la hormona paratiroidea en la enfermedad renal crónica. *Nefrología*. 2021;41:514-28.
31. Brown AJ, Ritter CS, Finch JL, Slatopolsky EA. Decreased calcium-sensing receptor expression in hyperplastic parathyroid glands of uremic rats: role of dietary phosphate. *Kidney Int*. 1999;55:1284-92.
32. Brown AJ, Dusso A, Lopez-Hilker S, Lewis-Finch J, Grooms P, Slatopolsky E. 1,25-(OH)₂D receptors are decreased in parathyroid glands from chronically uremic dogs. *Kidney Int*. 1989;35:19-23.
33. Dusso AS, Brown AJ, Slatopolsky E. Vitamin D. *Am J Physiol Renal Physiol*. 2005;289(1):F8-28.
34. Brown AJ, Zhong M, Finch J, Ritter C, McCracken R, Morrissey J, et al. Rat calcium-sensing receptor is regulated by vitamin D but not by calcium. *Am J Physiol*. 1996;270 3 Pt 2:F454-60.
35. Sherwood LM, Mayer GP, Ramberg CF Jr, Kronfeld DS, Aurbach GD, Potts JT Jr. Regulation of parathyroid hormone secretion: proportional control by calcium, lack of effect of phosphate. *Endocrinology*. 1968;83:1043-51.
36. Lopez-Hilker S, Dusso AS, Rapp NS, Martin KJ, Slatopolsky E. Phosphorus restriction reverses hyperparathyroidism in uremia independent of changes in calcium and calcitriol. *Am J Physiol*. 1990;259:F432-7.
37. Yi H, Fukagawa M, Yamato H, Kumagai M, Watanabe T, Kurokawa K. Prevention of enhanced parathyroid hormone secretion, synthesis and hyperplasia by mild dietary phosphorus restriction in early chronic renal failure in rats: possible direct role of phosphorus. *Nephron*. 1995;70:242-8.
38. Kilav R, Silver J, Naveh-Many T. Parathyroid hormone gene expression in hypophosphatemic rats. *J Clin Invest*. 1995;96:327-33.
39. Naveh-Many T, Sela-Brown A, Silver J. Protein-RNA interactions in the regulation of PTH gene expression by calcium and phosphate. *Nephrol Dial Transplant*. 1999;14:811-3.

40. Naveh-Many T. Minireview: the play of proteins on the parathyroid hormone messenger ribonucleic Acid regulates its expression. *Endocrinology*. 2010;151:1398-402.
41. Nechama M, Uchida T, Mor Yosef-Levi I, Silver J, Naveh-Many T. The peptidyl-prolyl isomerase Pin1 determines parathyroid hormone mRNA levels and stability in rat models of secondary hyperparathyroidism. *J Clin Invest*. 2009;119:3102-14.
42. Kilav-Levin R, Hassan A, Nechama M, Shilo V, Silver J, Ben-Dov IZ, et al. Post-transcriptional mechanisms regulating parathyroid hormone gene expression in secondary hyperparathyroidism. *FEBS J*. 2020;287:2903-13.
43. Hernández A, Salido E, Rodríguez M, Torres A. High phosphate diet increases prepro PTH ARNm independent of 1,25 (OH)₂D₃ and calcium in normal rats. *J Am Soc Nephrol*. 1995;6(963) (abstract).
44. Hernández A, Torres A, Concepción MT, Salido E. Parathyroid gland calcium receptor gene expression is not regulated by increased dietary phosphorus in normal and renal failure rats. *Nephrol Dial Transplant*. 1996;11 Suppl 3:11-4.
45. Hernández A, Concepción MT, Rodríguez M, Salido E, Torres A. High phosphorus diet increases preproPTH mRNA independent of calcium and calcitriol in normal rats. *Kidney Int*. 1996;50:1872-8.
46. Brown AJ, Zhong M, Ritter C, Brown EM, Slatopolsky E. Loss of calcium responsiveness in cultured bovine parathyroid cells is associated with decreased calcium receptor expression. *Biochem Biophys Res Commun*. 1995;212:861-7.
47. Almaden Y, Hernandez A, Torregrosa V, Canalejo A, Sabate L, Fernandez Cruz L, et al. High phosphate level directly stimulates parathyroid hormone secretion and synthesis by human parathyroid tissue in vitro. *J Am Soc Nephrol*. 1998;9:1845-52.
48. Estepa JC, Aguilera-Tejero E, Lopez I, Almaden Y, Rodriguez M, Felsenfeld AJ. Effect of phosphate on parathyroid hormone secretion in vivo. *J Bone Miner Res*. 1999;14:1848-54.
49. Almadén Y, Canalejo A, Ballesteros E, Añón G, Rodríguez M. Effect of high extracellular phosphate concentration on arachidonic acid production by parathyroid tissue in vitro. *J Am Soc Nephrol*. 2000;11:1712-8.
50. Almadén Y, Canalejo A, Ballesteros E, Añón G, Cañadillas S, Rodríguez M. Regulation of arachidonic acid production by intracellular calcium in parathyroid cells: effect of extracellular phosphate. *J Am Soc Nephrol*. 2002;13:693-8.
51. Canalejo A, Cañadillas S, Ballesteros E, Rodriguez M, Almaden Y. Importance of arachidonic acid as a mediator of parathyroid gland response. *Kidney Int Suppl*. 2003:S10-3.
52. de Francisco AL, Cobo MA, Setien MA, Rodrigo E, Fresnedo GF, Unzueta MT, et al. Effect of serum phosphate on parathyroid hormone secretion during hemodialysis. *Kidney Int*. 1998;54:2140-5.
53. Slatopolsky E, Finch J, Denda M, Ritter C, Zhong M, Dusso A, et al. Phosphorus restriction prevents parathyroid gland growth. High phosphorus directly stimulates PTH secretion in vitro. *J Clin Invest*. 1996;97:2534-40.
54. Denda M, Finch J, Slatopolsky E. Phosphorus accelerates the development of parathyroid hyperplasia and secondary hyperparathyroidism in rats with renal failure. *Am J Kidney Dis*. 1996;28:596-602.
55. Nielsen PK, Feldt-Rasmussen U, Olgaard K. A direct effect in vitro of phosphate on PTH release from bovine parathyroid tissue slices but not from dispersed parathyroid cells. *Nephrol Dial Transplant*. 1996;11:1762-8.
56. Almaden Y, Felsenfeld AJ, Rodríguez M, Cañadillas S, Luque F, Bas A, et al. Proliferation in hyperplastic human and normal rat parathyroid glands: role of phosphate, calcitriol, and gender. *Kidney Int*. 2003;64:2311-7.
57. Rodriguez M, Cañadillas S, Lopez I, Aguilera-Tejero E, Almaden Y. Regulation of parathyroid function in chronic renal failure. *J Bone Miner Metab*. 2006;24:164-8.
58. Lau WL, Obi Y, Kalantar-Zadeh K. Parathyroidectomy in the management of secondary hyperparathyroidism. *Clin J Am Soc Nephrol*. 2018;13(6):952-61.
59. Rodriguez ME, Almaden Y, Cañadillas S, Canalejo A, Siendones E, Lopez I, et al. The calcimimetic R-568 increases vitamin D receptor expression in rat parathyroid glands. *Am J Physiol Renal Physiol*. 2007;292(5):F1390-5.
60. Dusso AS, Pavlopoulos T, Naumovich L, Lu Y, Finch J, Brown AJ, et al. p21(WAF1) and transforming growth factor- α mediate dietary phosphate regulation of parathyroid cell growth. *Kidney Int*. 2001;59:855-65.
61. Arcidiacono MV, Sato T, Alvarez-Hernandez D, Yang J, Tokumoto M, Gonzalez-Suarez I, et al. EGFR activation increases parathyroid hyperplasia and calcitriol resistance in kidney disease. *J Am Soc Nephrol*. 2008;19:310-20.
62. Canalejo A, Canalejo R, Rodríguez ME, Martínez-Moreno JM, Felsenfeld AJ, Rodríguez M, et al. Development of parathyroid gland hyperplasia without uremia: role of dietary calcium and phosphate. *Nephrol Dial Transplant*. 2010;25:1087-97.
63. Shilo V, Ben-Dov IZ, Nechama M, Silver J, Naveh-Many T. Parathyroid-specific deletion of dicer-dependent microRNAs abrogates the response of the parathyroid to acute and chronic hypocalcemia and uremia. *FASEB J*. 2015;29:3964-76.
64. Hassan A, Levin R, Fisher Y, Silver J, Ben-Dov I, Naveh-Many T. The essential role of miRNA in maintaining an intact parathyroid in the adult. *Kidney Week*. 2021, virtual only, poster (PO0518) (abstract).
65. ADHR Consortium. Autosomal dominant hypophosphataemic rickets is associated with mutations in FGF23. *Nat Genet*. 2000;26:345-8.
66. Shimada T, Mizutani S, Muto T, Yoneya T, Hino R, Takeda S, et al. Cloning and characterization of FGF23 as a causative factor of tumor-induced osteomalacia. *Proc Natl Acad Sci U S A*. 2001;98:6500-5.
67. Richter B, Faul C. FGF23 actions on target tissues-with and without Klotho. *Front Endocrinol*. 2018;9:189.
68. Vervloet M. Fibroblast growth factor 23, the time is right for a second wind. *Kidney Int*. 2021;100:986-9.
69. Wolf M. Update on fibroblast growth factor 23 in chronic kidney disease. *Kidney Int*. 2012;82:737-47.
70. Gutiérrez OM. Fibroblast growth factor 23 and disordered vitamin D metabolism in chronic kidney disease: updating the "trade-off" hypothesis. *Clin J Am Soc Nephrol*. 2010;5:1710-6.
71. Lavi-Moshayoff V, Wasserman G, Meir T, Silver J, Naveh-Many T. PTH increases FGF23 gene expression and mediates the high-FGF23 levels of experimental kidney failure: a bone parathyroid feedback loop. *Am J Physiol Renal Physiol*. 2010;299:F882-9.
72. Llach F. Secondary hyperparathyroidism in renal failure: the trade-off hypothesis revisited. *Am J Kidney Dis*. 1995;25:663-79.
73. Komaba H, Fukagawa M. FGF23-parathyroid interaction: implications in chronic kidney disease. *Kidney Int*. 2010;77:292-8.
74. Kuro OM, Moe OW. FGF23- α Klotho as a paradigm for a kidney-bone network. *Bone*. 2017;100:4-18.
75. Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, et al. The parathyroid is a target organ for FGF23 in rats. *J Clin Invest*. 2007;117(12):4003-8.
76. Canalejo R, Canalejo A, Martínez-Moreno JM, Rodríguez-Ortiz ME, Estepa JC, Mendoza FJ, et al. FGF23 fails to inhibit uremic parathyroid glands. *J Am Soc Nephrol*. 2010;21:1125-35.
77. Navarro-García JA, Delgado C, Fernández-Velasco M, Val-Blasco A, Rodríguez-Sánchez E, Aceves-Ripoll J, et al. Fibroblast growth factor-23 promotes rhythm alterations and

- contractile dysfunction in adult ventricular cardiomyocytes. *Nephrol Dial Transplant*. 2019;34:1864–75.
78. Patel RB, Ning H, de Boer IH, Kestenbaum B, Lima JAC, Mehta R, et al. Fibroblast growth factor 23 and long-term cardiac function: the multi-ethnic study of atherosclerosis. *Circ Cardiovasc Imaging*. 2020;13:e011925.
79. Faul C, Amaral AP, Oskoueï B, Hu M-C, Sloan A, Isakova T, et al. FGF23 induces left ventricular hypertrophy. *J Clin Invest*. 2011;121:4393–408.
80. Scialla JJ, Wolf M. Roles of phosphate and fibroblast growth factor 23 in cardiovascular disease. *Nat Rev Nephrol*. 2014;10(5):268–78.
81. Navarro-González JF, Mora-Fernández C, Muros M, Herrera H, García J. Mineral metabolism and inflammation in chronic kidney disease patients: a cross-sectional study. *Clin J Am Soc Nephrol*. 2009;4:1646–54.
82. Muñoz Mendoza J, Isakova T, Ricardo AC, Xie H, Navaneethan SD, Anderson AH, et al. Fibroblast growth factor 23 and inflammation in CKD. *Clin J Am Soc Nephrol*. 2012;7:1155–62.
83. Izquierdo MC, Perez-Gomez MV, Sanchez-Niño MD, Sanz AB, Ruiz-Andres O, Poveda J, et al. Klotho, phosphate and inflammation/ageing in chronic kidney disease. *Nephrol Dial Transplant*. 2012;27 Suppl 4:iv6–10.
84. Cozzolino M, Elli F, Ciceri P, Ottaviano E, Conte F. Chapter 58 - Calcium and Phosphate Physiology. In: Ronco Claudio, Bellomo Rinaldo, Kellum John A, Ricci Zaccaria, editors. *Critical Care Nephrology (Third Edition)*. Elsevier; 2019. p. 345–9, e1, ISBN 9780323449427.
85. Almaden Y, Rodriguez-Ortiz ME, Canalejo A, Cañadillas S, Canalejo R, Martin D, et al. Calcimimetics normalize the phosphate-induced stimulation of PTH secretion in vivo and in vitro. *J Nephrol*. 2009;22:281–8.
86. Geng Y, Mosyak L, Kurinov I, Zuo H, Sturchler E, Cheng TC, et al. Structural mechanism of ligand activation in human calcium-sensing receptor. *Elife*. 2016:5.
87. Centeno PP, Herberger A, Mun H-C, Tu C, Nemeth EF, Chang W, et al. Phosphate acts directly on the calcium-sensing receptor to stimulate parathyroid hormone secretion. *Nat Commun*. 2019;10:4693.
88. Rodriguez M, Almaden Y, Hernandez A, Torres A. Effect of phosphate on the parathyroid gland: direct and indirect? *Curr Opin Nephrol Hypertens*. 1996;5:321–8.
89. Montes de Oca A, Madueño JA, Martínez-Moreno JM, Guerrero F, Muñoz-Castañeda J, Rodríguez-Ortiz ME, et al. High-phosphate-induced calcification is related to SM22 α promoter methylation in vascular smooth muscle cells. *J Bone Miner Res*. 2010;25:1996–2005.
90. Guerrero F, Herencia C, Almadén Y, Martínez-Moreno JM, Montes de Oca A, Rodríguez-Ortiz ME, et al. TGF- β prevents phosphate-induced osteogenesis through inhibition of BMP and Wnt/ β -catenin pathways. *PLoS One*. 2014;9:e89179.
91. Martínez-Moreno JM, Herencia C, de Oca AM, Díaz-Tocados JM, Vergara N, Gómez-Luna MJ, et al. High phosphate induces a pro-inflammatory response by vascular smooth muscle cells and modulation by vitamin D derivatives. *Clin Sci*. 2017;131:1449–63.
92. Muñoz-Castañeda JR, Herencia C, Pendón-Ruiz de Mier MV, Rodríguez-Ortiz ME, Díaz-Tocados JM, Vergara N, et al. Differential regulation of renal Klotho and FGFR1 in normal and uremic rats. *FASEB J*. 2017;31:3858–67.